

## SUPPLEMENTARY INFORMATION

### Pathogen profile

### *Pseudomonas savastanoi* pv. *savastanoi*: Some like it knot

Cayo Ramos<sup>1</sup>, Isabel M. Matas<sup>1</sup>, Leire Bardaji<sup>2</sup>, Isabel M. Aragón<sup>1</sup>, Jesús Murillo<sup>2\*</sup>

<sup>1</sup> Instituto de Hortofruticultura Subtropical y Mediterránea “La Mayora”, Universidad de Málaga-Consejo Superior de Investigaciones Científicas (IHSM-UMA-CSIC), Área de Genética, Facultad de Ciencias, Campus Teatinos s/n, E-29010 Málaga, Spain

<sup>2</sup> Departamento de Producción Agraria, ETS Ingenieros Agrónomos, Universidad Pública de Navarra, 31006 Pamplona, Spain

\* Corresponding author: Jesús Murillo; [jesus.murillo@unavarra.es](mailto:jesus.murillo@unavarra.es)

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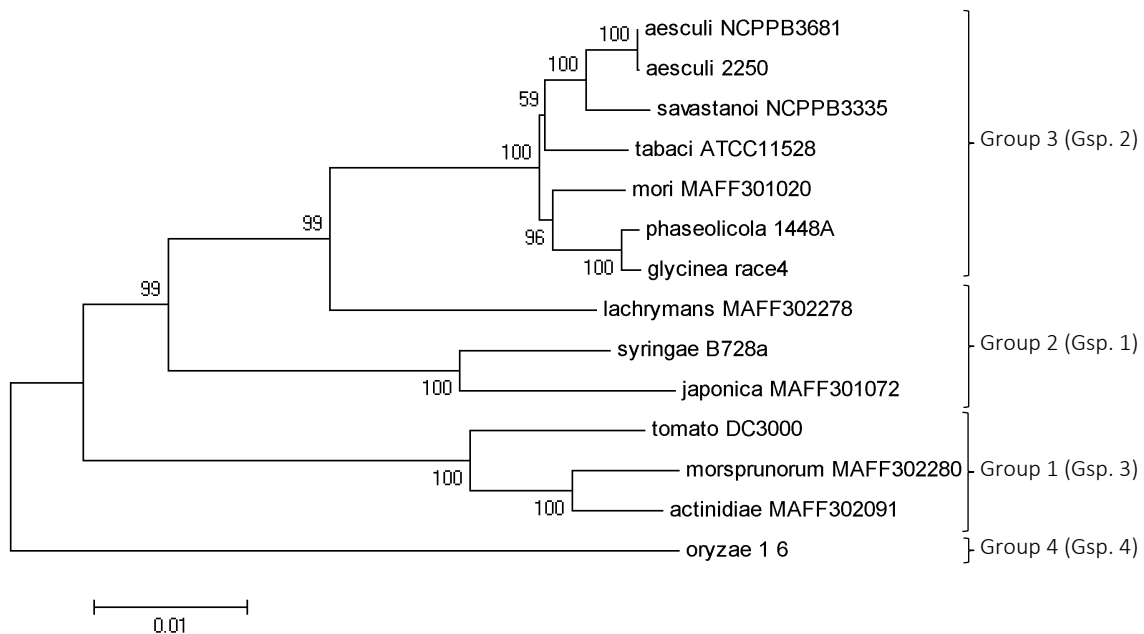
<http://onlinelibrary.wiley.com/doi/10.1111/j.1364-3703.2012.00816.x/supinfo>

**Fig. S1** Evolutionary relationships of *P. savastanoi* pv. *savastanoi* and selected *P. syringae* pathovars. Tree was constructed by multilocus sequence analysis using a concatenated data set (exactly 12000 nt) of *acnB*, *fruK*, *gapA*, *gltA*, *gyrB*, *pgi*, *recA* and *rpoD* genes. Phylogenetic groups 1, 2, 3 and 4 (Sarkar & Guttman, 2004, Studholme, 2011) correspond to genomospecies (Gsp) 3, 1, 2 and 4 (Gardan et al., 1999), respectively. Sequence alignment using Muscle, determination of the optimal nucleotide substitution model and phylogenetic tree construction were done using MEGA5 (Tamura et al., 2011); all positions containing gaps and missing data were eliminated using the option of complete deletion. Bootstrap values (1,000 repetitions) are shown on branches. Similar or identical topologies were obtained by maximum likelihood. The scale bar represents nucleotide substitutions per site.

**Table S1** Primers used for the detection of *Pseudomonas savastanoi* pv. *savastanoi*.

**Table S2** Comparison of the deduced products of *iaaM*-1 (PSA3335\_1475) and *iaaH*-1 (PSA3335\_1476), from *Pseudomonas savastanoi* pv. *savastanoi* NCPPB 3335, with their homologues in selected organisms.

**Table S3** Accession numbers and coordinates of the nucleotide sequences used for the construction of the neighbour-joining tree shown in Fig. 2.



**Fig. S1** Evolutionary relationships of *P. savastanoi* pv. *savastanoi* and selected *P. syringae* pathovars. Tree was constructed by multilocus sequence analysis using a concatenated data set (exactly 12000 nt) of *acnB*, *fruK*, *gapA*, *gltA*, *gyrB*, *pgi*, *recA* and *rpoD* genes. Phylogenetic groups 1, 2, 3 and 4 (Sarkar & Guttman, 2004, Studholme, 2011) correspond to genomospecies (Gsp) 3, 1, 2 and 4 (Gardan et al., 1999), respectively. Sequence alignment using Muscle, determination of the optimal nucleotide substitution model and phylogenetic tree construction were done using MEGA5 (Tamura *et al.*, 2011); all positions containing gaps and missing data were eliminated using the option of complete deletion. Bootstrap values (1,000 repetitions) are shown on branches. Similar or identical topologies were obtained by maximum likelihood. The scale bar represents nucleotide substitutions per site.

**Table S1** Primers used for the detection of *P. savastanoi* pv. *savastanoi*

Reference	Type of PCR	Target	Primer pair	Sequence (5' → 3')	Amplicon size (nt)	Observations
(Penyalver <i>et al.</i> , 2000, <i>Appl. Environ. Microbiol.</i> 66, 2673-2677)	Conventional	<i>iaaL</i> gene	IAALF IAALR	GGCACCAGCGGCAACATCAA CGCCCTCGGAACTGCCATAC	454	
(Bertolini <i>et al.</i> , 2003, <i>Phytopathology</i> , 93, 286-292)	Nested	<i>iaaL</i> gene	External pair IAALF-IAALR Internal pair IAALN1 IAALN2	See above  CTC CCT CTC CAA CGT CTT C GCC TGA TGA TTT TCT TCT G	338	Nested PCR in a single closed tube; sensitivity is highly increased by a dot blot hybridization step.
(Tegli <i>et al.</i> , 2010, <i>BMC Microbiol.</i> 10: 156-169)	Conventional	Chromosomal DNA; possible intergenic region	PsvF PsvR	GGCGATGTTCTCAGCGGATTTG GATCAAGTGTCCAAGGAAGTGAAGG	388	Authors designed primer pairs for the differential identification of pathovars <i>fraxini</i> , <i>nerii</i> and <i>savastanoi</i> .
	Real time		PsvRT-F PsvRT-R PsvRT-P	CGGATTTGTTTGCGGGGTA AATGGGGTGACACTAAAAATTGTGAA (HEX)CTCGTGCGATCTAAACAGCCGT AGC(BHQ-1)	298	

**Table S2** Comparison of the deduced products of *iaaM-1* (PSA3335\_1475) and *iaaH-1* (PSA3335\_1476), from *Pseudomonas savastanoi* pv. *savastanoi* NCPPB 3335, with their homologs in selected organisms<sup>a</sup>

Strain	<i>iaaM-1</i>			<i>iaaH-1</i>		
	Prot. Acc. number	Id.	Similarity	Prot. Acc. number	Id.	Similarity
<i>P. savastanoi</i> pv. <i>nerii</i> EW2009	AAA25852	100.0	100.0	AAA25853	97.6	97.6
<i>P. syringae</i> pv. <i>syringae</i> B728a	AAY36585	94.4	97.7	AAY36586	92.6	95.5
<i>P. savastanoi</i> pv. <i>savastanoi</i> NCPPB 3335 ( <i>iaaM-2</i> and <i>iaaH-2</i> )	(-) <sup>b</sup>			EFI00888	92.4	95.3
<i>P. syringae</i> pv. <i>aceris</i> MAFF302273	EGH68944	94.4	97.7	EGH68943	92.2	95.5
<i>P. syringae</i> pv. <i>glycinea</i> race 4	AEGH01000077 (1829-3502) <sup>c</sup>	91.9	95.0	EFW83486	91.9	94.6
<i>P. syringae</i> pv. <i>glycinea</i> B076	AEGG01000022 (403-2076) <sup>c</sup>	91.9	95.0	EFW81447	91.9	94.6
<i>Dickeya dadantii</i> 3937	ADM96599	66.2	77.4	ADM96600	58.1	73.1
<i>Burkholderia phymatum</i> STM815	ACC76653	69.0	79.2	ACC76652	55.6	72.6
<i>Pantoea agglomerans</i> pv. <i>gypsophilae</i> PD713	AAC17187	69.1	80.1	AAC17186	55.2	68.7
<i>P. chlororaphis</i> subsp. <i>chlororaphis</i> GP72	AHAY01000223 (4470-6176) <sup>c</sup>	55.8	72.5	AHAY01000223 (4430-3102) <sup>c</sup>	42.7	57.6
<i>Agrobacterium tumefaciens</i> C58	AAD30489	36.8	50.9	AAD30488	30.1	45.3

<sup>a</sup> Global alignments were done using the EMBOSS Needle Protein Sequence Alignment interface([http://www.ebi.ac.uk/Tools/psa/emboss\\_needle/](http://www.ebi.ac.uk/Tools/psa/emboss_needle/)); figures represent percentage of amino acid identity (Id.) and similarity.<sup>b</sup> *iaaM-2* (PSA3335\_1017) is a pseudogene.<sup>c</sup> In these cases, proteins were deduced from the corresponding nucleotide contig sequences, with figures in parenthesis indicating the CDS coordinates.

**Table S3** Accession numbers and coordinates of the nucleotide sequences used for the construction of the NJ tree shown in Figure 2

Strain	Gsp. <sup>a</sup>	Accession number	Coordinates <sup>b</sup>
<i>P. cannabina</i> pv.			
<i>alisalensis</i> ES4326 <sup>c</sup>	9	AEAK01000502	41622-42809
<i>P. syringae</i> pv.			
<i>actinidiae</i> CRA-FRU 8.43	8	AFTG01000160	3312-4499
<i>actinidiae</i> MAFF302091	8	AEAL01000291	3491-4678
<i>actinidiae</i> NCPPB 3739	8	AFTH01000772	5774-6961
<i>actinidiae</i> NCPPB 3871	8	AFTF01000420	5775-6962
<i>aesculi</i> 2250	2	ACXT01000531	14852-16039
<i>aesculi</i> NCPPB 3681 (0893_23) <sup>d</sup>	2	ACXS01000384	3051-4238
<i>glycinea</i> B076	2	AEGG01000003	5442-6577
<i>glycinea</i> race 4	2	ADWY01001392	1-1133
<i>lachrymans</i> MAFF302278	3	AEAM01000068	15292-16479
<i>mori</i> MAFF301020	2	AEAG01001010	58-1245
<i>morsprunorum</i> MAFF302280	uncertain	AEAE01000140	20038-21225
<i>oryzae</i> 1_6	4	ABZR01000421	2353-3540
<i>tabaci</i> ATCC 11528	2	ACHU02000160	9306-10493
<i>theae</i> NCPPB 2598	8	AGNN01000139	1477-2664
tomato DC3000	3	NC_004578	406323-407510
tomato K40	3	ADFY01000268	14995-16182
tomato Max13	3	ADFZ01000040	39074-40261
tomato NCPPB 1108	3	ADGA01000161	20655-21842
tomato T1	3	ABSM01000063	2149-3318
<i>P. savastanoi</i> pv.			
<i>nerii</i> EW2009	2	M35373	1100-2287
<i>savastanoi</i> NCPPB 3335 <i>iaaL</i> 1	2	GG774623	112-1286
<i>savastanoi</i> NCPPB 3335 <i>iaaL</i> 2	2	NZ_GG774681	14330-13137

<sup>a</sup> Gsp, genomospecies; data taken from Gardan et al. (1999 Int. J. Syst. Bacteriol. 49,469-478) and Parkinson et al. (2011 Plant Pathol. 60,338-344).

<sup>b</sup> Position of the *iaaL* CDS within the sequence.

<sup>c</sup> *P. cannabina* pv. *alisalensis* ES4326 was previously designated *P. syringae* pv. *maculicola* ES4326 (Bull et al. 2011 Phytopathology 101,847-858).

<sup>d</sup> These names were reported as synonymous (Green et al. 2010 PLoS ONE 5(4):e10224)