



## OPEN ACCESS

## EDITED BY

Litao Yang,  
Shanghai Jiao Tong University, China

## REVIEWED BY

Antonio Lupini,  
Mediterranea University of Reggio Calabria,  
Italy  
Litao Yang,  
Shanghai Jiao Tong University, China  
Jianxin Shi,  
Shanghai Jiao Tong University, China

## \*CORRESPONDENCE

Sergio G. Nebauer  
✉ sergonne@bvg.upv.es  
Joaquín Medina  
✉ medina.joaquin@inia.csic.es†These authors have contributed equally to  
this work

## SPECIALTY SECTION

This article was submitted to  
Plant Biotechnology,  
a section of the journal  
Frontiers in Plant Science

RECEIVED 03 August 2022

ACCEPTED 13 February 2023

PUBLISHED 01 March 2023

## CITATION

Carrillo L, Baroja-Fernández E,  
Renau-Morata B, Muñoz FJ, Canales J,  
Ciordia S, Yang L, Sánchez-López AM,  
Nebauer SG, Ceballos MG,  
Vicente-Carbajosa J, Molina RV,  
Pozueta-Romero J and Medina J (2023)  
Ectopic expression of the *AtCDF1*  
transcription factor in potato enhances  
tuber starch and amino acid contents and  
yield under open field conditions.  
*Front. Plant Sci.* 14:1010669.  
doi: 10.3389/fpls.2023.1010669

## COPYRIGHT

© 2023 Carrillo, Baroja-Fernández, Renau-  
Morata, Muñoz, Canales, Ciordia, Yang,  
Sánchez-López, Nebauer, Ceballos, Vicente-  
Carbajosa, Molina, Pozueta-Romero and  
Medina. This is an open-access article  
distributed under the terms of the [Creative  
Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The  
use, distribution or reproduction in other  
forums is permitted, provided the original  
author(s) and the copyright owner(s) are  
credited and that the original publication in  
this journal is cited, in accordance with  
accepted academic practice. No use,  
distribution or reproduction is permitted  
which does not comply with these terms.

# Ectopic expression of the *AtCDF1* transcription factor in potato enhances tuber starch and amino acid contents and yield under open field conditions

Laura Carrillo<sup>1†</sup>, Edurne Baroja-Fernández<sup>2†</sup>,  
Begoña Renau-Morata<sup>3†</sup>, Francisco J. Muñoz<sup>2</sup>,  
Javier Canales<sup>4,5</sup>, Sergio Ciordia<sup>6</sup>, Lu Yang<sup>1</sup>,  
Ángela María Sánchez-López<sup>2</sup>, Sergio G. Nebauer<sup>7\*</sup>,  
Mar G. Ceballos<sup>1</sup>, Jesús Vicente-Carbajosa<sup>1</sup>, Rosa V. Molina<sup>7</sup>,  
Javier Pozueta-Romero<sup>8</sup> and Joaquín Medina<sup>1\*</sup><sup>1</sup>Centro de Biotecnología y Genómica de Plantas (CBGP) UPM-INIA/CSIC, Campus de Montegancedo, Madrid, Spain, <sup>2</sup>Instituto de Agrobiotecnología (IdAB), CSIC-Gobierno de Navarra, Mutiloabeti, Nafarroa, Spain, <sup>3</sup>Departamento de Biología Vegetal, Universitat de València. Vicent Andrés Estellés, Burjassot, Spain, <sup>4</sup>Instituto de Bioquímica y Microbiología, Facultad de Ciencias, Universidad Austral de Chile, Valdivia, Chile, <sup>5</sup>ANID–Millennium Science Initiative Program, Millennium Institute for Integrative Biology (iBio), Santiago, Chile, <sup>6</sup>Unidad Proteómica (CNB), Centro Nacional de Biotecnología (CNB–CSIC), Cantoblanco, Madrid, Spain, <sup>7</sup>Departamento de Producción Vegetal, Universitat Politècnica de València, València, Spain, <sup>8</sup>Institute for Mediterranean and Subtropical Horticulture “La Mayora” (IHSM), CSIC-UMA, Málaga, Spain**Introduction:** Cycling Dof transcription factors (CDFs) have been involved in different aspects of plant growth and development. In Arabidopsis and tomato, one member of this family (CDF1) has recently been associated with the regulation of primary metabolism and abiotic stress responses, but their roles in crop production under open field conditions remain unknown.**Methods:** In this study, we compared the growth, and tuber yield and composition of plants ectopically expressing the *CDF1* gene from Arabidopsis under the control of the 35S promoter with wild-type (WT) potato plants cultured in growth chamber and open field conditions.**Results:** In growth chambers, the *35S::AtCDF1* plants showed a greater tuber yield than the WT by increasing the biomass partition for tuber development. Under field conditions, the ectopic expression of *CDF1* also promoted the sink strength of the tubers, since *35S::AtCDF1* plants exhibited significant increases in tuber size and weight resulting in higher tuber yield. A metabolomic analysis revealed that tubers of *35S::AtCDF1* plants cultured under open field conditions accumulated higher levels of glucose, starch and amino acids than WT tubers. A comparative proteomic analysis of tubers of *35S::AtCDF1* and WT plants cultured

under open field conditions revealed that these changes can be accounted for changes in the expression of proteins involved in energy production and different aspects of C and N metabolism.

**Discussion:** The results from this study advance our collective understanding of the role of CDFs and are of great interest for the purposes of improving the yield and breeding of crop plants.

#### KEYWORDS

potato, CDF, crop yield, photoassimilate partition, amino acid, starch, C/N metabolism

## Introduction

Potato is now the fourth most important food crop worldwide, after wheat, rice and corn and the major staple food in many developing countries (FAO, 2020). In addition, the potato is also grown for processed food, starch production or animal feed. Their tubers are rich in starch (60–80% of the dry matter) as well as quality proteins, vitamins, micronutrients and antioxidants (Burgos et al., 2020). As a major food crop, the potato has a key role in the 2030 agenda for sustainable development, and is vital if the goals aimed at ensuring nutritious and sufficient food are to be met ([www.faostat.fao.org](http://www.faostat.fao.org)).

In the current context of climate change, where a substantial decline in potato yield has been predicted (Haverkort and Struik, 2015; Dahal et al., 2019), improving the production and nutritional quality of tubers is key if agricultural development is to take place (Hameed et al., 2018). Therefore, despite the rise in global production scale during the last few decades, increasing the yield per hectare is a crucial breeding objective in the case of potato (Bradshaw, 2021). The aim behind conventional breeding techniques has been directed to increase yield, processing and storage quality, as well as to enhance disease resistance (Birch et al., 2012). Although these strategies have been successfully employed for the introduction of some traits, such as the resistance to fungal pathogens (Naess et al., 2000; Tek et al., 2004; Halterman et al., 2016), the progress is slow, and it is a challenging task due to tetraploidy, heterozygosity and a lack of variability as regards the traits of interest in commercial cultivars (Patil et al., 2016). Wild species represent a valuable germplasm source, but some wild and cultivated species exhibit sexual incompatibility (Halterman et al., 2016). In this context, during the last few years, the efforts to improve this crop have focused on the development of biotech potato varieties (Barrell et al., 2013). In addition to the improvement of tolerance to pathogens (Martínez-Prada et al., 2021), several analyses succeeded in incorporating nutritional traits in potato, such as enhanced protein content (Chakraborty et al., 2010) and increased levels of some essential amino acids (Dancs et al., 2008; Kumar and Jander, 2017), vitamin C (Bulley et al., 2012),  $\beta$ -carotene (Li et al., 2012) or triacylglycerols (Hofvander et al., 2016), as well as a higher amylose content (Krunić et al., 2018). Furthermore, the reduction of anti-nutrient

contents in potato, such as steroidal glycoalkaloids, has also been addressed (Itkin et al., 2013). It is noteworthy that in some of these analyses (Dancs et al., 2008), yield reductions of up to 60% have been described. Only a few reports have shown that genetic engineering can produce transgenic events with an improved tuber yield greater than that produced by conventional breeding under field conditions (Baroja-Fernández et al., 2009; Pourazari et al., 2018). As pointed out by Pourazari et al. (2018), breeding for improved crop quality traits can affect non-target traits related to growth and resource use, and it is important to take into account that these effects may vary in different cultivation conditions (e.g., greenhouse vs. field). Therefore, the selection of suitable genomic targets able to change metabolic aspects while increasing the yield under agronomic crop management conditions is a critical prerequisite if the desired goals are to be reached. In this scenario, we have surveyed the use of DOF transcription factors (like CDFs), which has been associated with the regulation of the primary metabolism in other *Solanaceae* species (Renau-Morata et al., 2017; Domínguez-Figueroa et al., 2020) as a new genetic tool to increase crop production and nutritional quality.

Plant-specific DNA binding with one finger (DOF) proteins are a class of transcription factors (TFs) that exhibit a 50 amino acid conserved DNA binding domain that binds to the DNA motif: 5'-T/AAAG-3' (Yanagisawa and Schmidt, 1999). DOF proteins play different functions in plant growth and development comprising root growth, shoot branching, seed and tuber development, and flowering (Yanagisawa, 2002; Kloosterman et al., 2013; Noguero et al., 2013; Rueda-Lopez et al., 2017). Maize *ZmDOF1* and *ZmDOF2* have also been involved in C/N balance and nitrogen assimilation (Yanagisawa and Sheen, 1998; Yanagisawa, 2004; Peña et al., 2017) by the differential control of the expression of central genes like *pyruvate kinase (PK)*, *phosphoenolpyruvate carboxylase (PEPC)* and *glutamate synthase (GS)*. In fact, the ectopic expression of *ZmDOF1* in both rice and *Arabidopsis* increased the expression levels of key genes involved in carbon skeleton production (e.g. *C4-PEPC* and *PK1*) and N assimilation (e.g. *GS* and *glutamate synthase, GOGAT*) (Yanagisawa, 2004; Kurai et al., 2011; Peña et al., 2017).

Amongst different DOFs, cycling DOF Factors (CDFs) play key functions in the photoperiodic pathway, modulating flowering time by controlling the expression of *CONSTANS (CO)* and *FLOWERING LOCUS (FT)* (Imaizumi et al., 2005; Fornara et al.,

2009; Corrales et al., 2014). In long days, CDF protein stability is tightly controlled by a protein complex constituted by the protein encoded by clock gene *GIGANTEA (GI)* (Park et al., 1999) and the blue-light receptor *FLAVIN BINDING KELCH REPEAT F-BOX PROTEIN 1 (FKF1)*, through specific conserved motifs of 10–30 amino acid residues, located in the CDF C-terminal region (Imaizumi et al., 2005; Sawa et al., 2007; Kloosterman et al., 2013). In potato, it has been described that a *StCDF1* is involved in the photoperiodic control of tuberization by modulating the expression of StSP6A mobile tuberization signal in a StGI and StFKF1 dependent manner (Kloosterman et al., 2013; Abelenda et al., 2019). Potato plants expressing the alleles *StCDF1.2* and *StCDF1.3*, which code for truncated forms lacking the StFKF1 binding domain, are long-day dependent for tuberization, in sharp contrast, genotypes expressing the allele *StCDF1.1* encoding a full-length protein are strictly short-day dependent for tuberization (Kloosterman et al., 2013).

CDFs have also been associated with abiotic stress responses (Fornara et al., 2009; Corrales et al., 2017; Renau-Morata et al., 2017; Renau-Morata et al., 2020a). Arabidopsis *AtCDFs* and tomato *SICDFs* genes respond to abiotic stresses including dehydration, and osmotic, heat and cold stress (Corrales et al., 2014; Corrales et al., 2017). The ectopic expression of tomato *SICDF1* and *SICDF3* genes in Arabidopsis improved tolerance to drought and salinity (Corrales et al., 2014). Moreover, the overexpression of *SICDF3* in tomato enhanced salt tolerance (Renau-Morata et al., 2017). In the case of potato, the *StCDF1* gene and the long non-coding RNA (lncRNA) counterpart (FLORE), are involved in the control of water loss by modulating both stomatal growth and opening control (Ramírez-Gonzales et al., 2021).

Recent studies have provided evidence for the possible involvement of CDFs in the regulation of nitrogen and carbon metabolisms. Network analyses of a group of 2,174 dynamic N-responsive genes allowed the identification of CDF1 as a key TF that is connected with a second layer of TFs in Arabidopsis (Varala et al., 2018). Using the TARGET system, these studies showed that the nitrogen master factor NLP7 binds the promoter region of CDF1, suggesting that CDF1 is a component of the regulatory network involved in Nitrate-signalling cascade (Alvarez et al., 2020). In addition, in Arabidopsis and tomato, we showed that the ectopic expression of *AtCDF3*, the closest homolog of *AtCDF1*, promoted significant changes in the primary metabolism in leaves, including increased levels of sucrose, glutamine, asparagine and GABA in leaves (Corrales et al., 2017; Renau-Morata et al., 2017). Moreover, the ectopic expression of *AtCDF3* in tomato modified the fruit's amino acid and sugar contents and resulted in enhanced yield and nutrient use efficiency (NUE) under greenhouse conditions (Renau-Morata et al., 2017; Domínguez-Figueroa et al., 2020). Additional functions of tomato *SICDFs* in carbon partitioning and carbon metabolism have been described (Renau-Morata et al., 2020a). The targeted overexpression of *SICDF4* gene in the tomato fruit increased fruit size and total plant yield and promoted a greater partition of carbon resources to the fruit which was related to more sink strength and an increased activity of the sucrose synthase and starch biosynthetic enzymes involved in the sucrose-to-starch conversion process (Renau-Morata et al., 2020b). The overall data

indicated that CDFs play significant roles in plant growth and C and N metabolism, and the partition of photoassimilates to the main developing sinks. Therefore, these genes might be used as tools to improve crop yield.

Herein, we have explored the effects of ectopic *AtCDF1* expression in tuber composition and the yield and growth of potato plants cultured under both growth chamber and open field conditions. Notably, a higher tuber yield was obtained in the 35S:: *AtCDF1* potato plants under agronomic management field conditions, as also observed in growth chambers. The higher production of tubers could be explained by an increased partition of photoassimilates to the developing tubers, leading to a higher proportion of bigger tubers corresponding to the consumption size-category. In addition, also changes in the contents of starch, sugars and quality amino acids, all of which are related to improved tuber quality, were also observed in metabolite analyses. We also characterized changes at the molecular level promoted by the ectopic expression of *AtCDF1* and showed that the overexpression of *AtCDF1* in potato increases tuber yield and improves tuber composition through specific mechanisms involving proteome resetting. Together, our results confirmed the potential of CDF manipulation in breeding programs for crop yield and quality, since a greater sink strength is promoted in the main harvested sink, irrespective of the type of organ.

## Materials and methods

### Potato transformation and growth conditions

The ORFs of the *AtCDF1* gene (*Arabidopsis thaliana*, Col-0) genes were amplified by PCR using cDNA as a template and cloned into a pROK2 vector under the control of CaMV 35S promoter, and the nopaline synthase gene (*NOS*) terminator. The *Agrobacterium tumefaciens* strain LBA4404, was transformed with the resultant plasmid to transform Desiree variety potato plants following the method described by Kiel et al. (1989) with several modifications. Potato transgenic plants were generated by co-culturing leave explants from 4-week-old plants maintained *in vitro* and *A. tumefaciens* harbouring *AtCDF1* construct. To prepare the explants, the petiole and ¼ of the leaf in immediate proximity to the petiole were discarded. The middle of the remaining leaf was manually wounded in two or three places to permit the entry of *Agrobacteria* into the cells and intercellular spaces. The explant size was approximately of 1 cm<sup>2</sup>. Twenty independent explants were placed in a Petri dish with 40 mL of MS liquid medium (MS medium with salts and vitamins, 20 g L<sup>-1</sup> sucrose, 0.5 g L<sup>-1</sup> MES, pH=5.7) containing 320 µL of an *A. tumefaciens* culture (DO<sub>600 nm</sub>=0,8), and incubated at 22°C and without light for two days. Sixty explants were used for each construction to be transformed. Next, the explants were dried on filter paper and cultured on callus induction medium (CIM; 4.4 g L<sup>-1</sup> MS salts, 16 g L<sup>-1</sup> 6-Benzylaminopurine (BAP), 5 mg L<sup>-1</sup> α-naphthaleneacetic acid (NAA), 7 g L<sup>-1</sup> agar, 50 mg L<sup>-1</sup> kanamycin, 250 mg L<sup>-1</sup> cefotaxime) for 7 days. Subsequently, the explants were transferred to shoot induction medium (SIM; 4.4 g L<sup>-1</sup> MS salts, 16 g L<sup>-1</sup> glucose, 0.5 g L<sup>-1</sup> MES, 0.02

mg L<sup>-1</sup> (NAA), 0.02 mg L<sup>-1</sup> gibberellic acid A<sub>3</sub> (GA<sub>3</sub>), 2 mg L<sup>-1</sup> zeatin riboside, 7 g L<sup>-1</sup> agar, 50 mg L<sup>-1</sup> kanamycin, 250 mg L<sup>-1</sup> cefotaxime). During shoot generation, the explants were regularly transferred to a fresh medium in 7-10 days intervals until shoots were regenerated. Individual shoots reaching a length of 15-20 mm were excised and transferred to root induction medium (RIM; 4.4 g L<sup>-1</sup> MS salts, 16 g L<sup>-1</sup> glucose, 0.5 g L<sup>-1</sup> MES, 7 g L<sup>-1</sup> agar, 50 mg L<sup>-1</sup> kanamycin, 250 mg L<sup>-1</sup> cefotaxime). Plantlets with well-developed roots were multiplied in *in vitro* culture or transferred to soil for growth chamber and field experiments.

For the assays in growth chambers, the *AtCDF1* overexpressor and WT potato (cv. Desiree) plants were grown under a long day photoperiod of 16-h light/8-h dark photoperiod and a thermoperiod of 23/18°C (light/dark). Plants were grown in 0.5 L pots were fertilized with half-strength Hoagland n°2 solution (Hoagland and Arnon, 1950) for 45 days, then, plant biomass and photosynthesis parameters were determined.

## RNA extraction and qPCR analyses

Total RNA was extracted from leaves and tubers of 2-week-old and 3-month-old potato plants, respectively, following the protocol of Oñate-Sánchez and Vicente-Carvajosa (2008) and treated with DNase (Roche). Then, cDNA was synthesized using 2 µg of DNA-free RNA and the avian myeloblastosis virus reverse transcriptase and oligo-(dT)15 primers (Promega) according to the manufacturer's instructions.

The expression level of *AtCDF1* gene in overexpression lines was determined by RT-qPCR. A LightCycler®480 System (Roche) was used for real-time PCR (10 s at 95°C, 45 cycles of 95°C for 10 s, 60°C for 20 s min and 72°C for 30 s) using LightCycler® 480 SYBR Green I Master (Roche). The *UBIQUITIN* gene from *S. tuberosum* (PGSC0003DMG400004605) was used as reference gene to normalize all measurements. The oligonucleotides used are: 5'-TGCTTACCGTCTCGTCTTCG-3' and 5'-TCCGGGAGT TTCGTGGCCGT-3' for *AtCDF1*. 5'-AGAAGGCCGG GTGCGTTCTG-3' and 5'-ACCGGCTTGCACATCGCCT-3' for *UBIQUITIN* (PGSC0003DMG400004605). The relative expression levels of the target genes were calculated using the 2<sup>-ΔΔCT</sup> method (Livak and Schmittgen, 2001).

## Measurement of photosynthetic activity

Instantaneous values of the net CO<sub>2</sub> assimilation rate (A<sub>n</sub>), stomatal conductance (g<sub>s</sub>), substomatal CO<sub>2</sub> concentration (C<sub>i</sub>) and transpiration rate (E) were determined with an LI-6400 infrared gas analyser (LICOR Biosciences, USA). The conditions in the measuring chamber were controlled at a saturating PAR of 1,000 µmol m<sup>-2</sup> s<sup>-1</sup>, 400 ppm CO<sub>2</sub> and relative humidity of 60-70%. The attached leaf chamber fluorometer enabled the measurement of the effective quantum yield of PSII (PhiPS2). One measurement per plant was taken on the third or fourth leaf from the apex. Eight to ten different plants of each genotype were used.

## Experiment under field conditions

Field trials were conducted in Tudela (Navarra, Spain) during May and September, 2016. Two independent 35S::*AtCDF1* lines (L5 and L6), and WT potato (cv. Desiree) plants were used in the field assays. For agronomic analyses, the plants were randomly distributed in three 50 m<sup>2</sup> parcels, each one containing 30 plants per line. The plants were grown in a 35 cm spacing plant-plant (distance) in rows 90-cm apart. After 5 months of growth, at the end of the growing season, the total tuber fresh weight (Kg tubers/ha) and total tuber number (no of tubers/ha) were measured. The tubers were graded into three tuber size classes: 0-25 mm (out grade), 25-40 mm (seed) and greater than 40 mm (consumption) and the tuber fresh weight and number per size were measured. The total yield included all the collected tubers (out grades, greens, mechanically damaged, misshapen) while the commercial yield included only seed and consumption sizes. The standard management practices for potato (fertilization and pest/pathogen control) were conducted.

## Determination of the sugar and protein content, amino acid composition and energy-related parameters

For the measurement of the sugar content, total protein content, amino acid composition and energy-related parameters tubers of 30-50 g FW, were harvested from senescent plants, and used. The tuber samples were obtained as described in Baroja-Fernández et al. (2009). Three samples were taken from each tuber, immediately freeze-clamped and finally ground to a fine powder in liquid nitrogen.

Soluble sugars (glucose, fructose and sucrose) were determined by HPLC on an ICS-3000 Dionex system as described by Baroja-Fernández et al. (2003). The protein content was determined by using Bradford assay reagent (Bio-Rad XL-100). The starch content was measured enzymatically with an amyloglucosidase-based UV-test Kit (R-Biopharm).

The amylose content in tuber starch was determined using the iodine-based colorimetric method (Andersson et al., 2006). Adenine nucleotides were extracted using HClO<sub>4</sub> and measured by HPLC with a Partisil 10-SAX column according to Bahaji et al. (2015). The measurement of reduced (NADH, NADPH) and oxidized nicotinamide adenine dinucleotides (NAD<sup>+</sup> and NADP<sup>+</sup>) were measured as described by Queval and Noctor (2007). The amino acids profile was analyzed using HPLC, as described in Loiret et al. (2009).

## Protein digestion for proteomic analyses

For the proteomic analyses tubers of 30-50 g FW, harvested from senescent plants in the field trial, were used. The samples were resuspended in lysis buffer containing 8.4 M urea, 2.4 M thiourea, 5% CHAPS, 5 mM TCEP, and a protease inhibitor cocktail (Merck)

and incubated for 15 min on ice. The pellet was homogenized by ultrasonication for 5 min on ultrasonic bath, Branson 2510 (Marshall Scientific), was centrifuged at  $20000 \times g$  for 10 min at  $4^{\circ}\text{C}$ , and the supernatant containing the solubilized proteins was used for further analysis. Then, 20  $\mu\text{g}$  of protein was precipitated by methanol/chloroform method and resuspended in 20  $\mu\text{L}$  of multichaotropic sample solution UTT buffer (7 M Urea, 2 M thiourea, 100 mM TEAB; Merck).

The sample was then reduced-alkylated with 2  $\mu\text{L}$  of 50 mM TCEP, pH 8.0, at  $37^{\circ}\text{C}$  for 60 min and 1  $\mu\text{L}$  of 200 mM cysteine-blocking reagent MMTS (SCIEX, Foster City, CA) for 10 min at room temperature. The sample was diluted to 140  $\mu\text{L}$  with 25 mM TEAB and the digestion was initiated by adding 1  $\mu\text{g}$  of Pierce MS-grade trypsin (Thermo-Fisher Scientific Inc.) to each sample in a ratio of 1:20 (w/w) and then incubated at  $37^{\circ}\text{C}$  overnight. The sample digestion was evaporated to dryness in a vacuum concentrator.

## Liquid chromatography and mass spectrometer analysis

The digested sample was cleaned up/desalted using Stage-Tips C18 (Merck) and the peptide concentration was determined by Qubit<sup>TM</sup> Fluorometric Quantitation (Thermo Fisher Scientific). A 1  $\mu\text{g}$  aliquot of each digested sample was subjected to 1D-nano LC-ESI-MS/MS analysis using a nano liquid chromatography system (Eksigent Technologies nanoLC Ultra 1D plus, SCIEX) coupled to a high-speed TripleTOF 5600 mass spectrometer (SCIEX) with a Nanospray III source. Peptides were separated using a 250 min gradient ranging from 2% to 90% mobile phase B (mobile phase A: 2% acetonitrile, 0.1% formic acid; mobile phase B: 100% acetonitrile, 0.1% formic acid).

The data were acquired using an ion spray voltage floating (ISVF) 2300 V, curtain gas (CUR) 35, interface heater temperature (IHT) 150, ion source gas 1 (GS1) 25, and declustering potential (DP) 100 V. All the data were acquired using information-dependent acquisition (IDA) mode (0.25 s MS survey scan in the mass range of 350–1250 Da and 35 MS/MS scans of 100 ms in the mass range of 100–1800 Da) using Analyst TF 1.7 software (SCIEX, USA).

## Data analysis and quantification

The raw instrument files were processed and exported as mgf files. The proteomics data analysis was performed by using 4 search engines (Mascot Server v.2.6.1, OMSSA, XTandem, and Myrimatch) and a target/decoy database built from sequences in the *S. tuberosum* proteome at Uniprot Knowledgebase.

All the searches were configured with dynamic modifications for pyrrolidone from Q (-17.027 Da) and the oxidation of methionine residues (+15.9949 Da) and static modification such as methylthiolation (+45.987 Da) on cysteine, monoisotopic masses, and trypsin cleavage (max 2 missed cleavages). The peptide precursor mass tolerance was 10 ppm, and the MS/MS tolerance was 0.02 Da. Score distribution models were used to

compute peptide-spectrum match p-values (Ramos-Fernández et al., 2008), and the spectra recovered by an  $\text{FDR}^* \leq 0.01$  (peptide-level) filter were selected for quantitative analysis. The differential regulation was measured using linear models (Lopez-Serra et al., 2014), and statistical significance was measured using q-values (FDR, False Discovery Rate). All the analyses were conducted using software from Proteobotics (Madrid, Spain). The data were deposited at ProteomeXchange Consortium PXD036489. The Gene Ontology (GO) enrichment analyses were performed on the up- and down-regulated proteins using gPROFILER software (Reimand et al., 2016). Significantly enriched ( $\text{Padj} < 0.05$ ) GO terms describing a biological process (GO BP) for the differential expressed proteins are displayed shown in the figures. The significant metabolic pathways were analyzed using the Mapman tool (Schwacke et al., 2019). The most over-represented biological functions are shown in Table S2.

## Data analysis and statistics

The data sets were subjected to analysis of variance (ANOVA) procedures with the Statgraphics statistical software (Statgraphics Centurion XVI, Statpoint Tech, Inc., USA). The mean values were analyzed ( $p < 0.05$  or  $p < 0.01$ ) by a Student-Newman-Keuls test.

## Results

### The ectopic expression of *AtCDF1* increases the potato tuber yield

In order to examine the functions of *AtCDF1* and to assess its impact on plant growth and production in potato, a detailed phenotypic characterization of *AtCDF1* gain-of-function plants was conducted by analyzing their performance. Thus, three independent *35S::AtCDF1* lines (L2, L5 and L6) were obtained as described in Materials and Methods. As shown in Figure S1, *AtCDF1* transcript levels in leaves and tubers were much higher in all the transgenic *35S::AtCDF1* lines than in the WT plant biomass and photosynthesis were measured in the *35S::AtCDF1* and WT potato plants after 45 days of growth at  $24^{\circ}\text{C}$  under long-day photoperiod (16h light/8 h dark) in growth chamber conditions. As shown in Figures 1, S1, S2, the transgenic plants and WT showed similar external phenotypes as well as total plant biomass (Figure S2E). Accordingly, there were no significant differences photosynthesis activity of WT and *35S::AtCDF1* plants, since similar net photosynthetic rates and effective quantum yields of PSII were observed (Figures S1A, D). Notably, *35S::AtCDF1* potato plants (L2, L5 and L6 lines) produced 105–136% more tubers (Figure 1B) and a 41–56% higher total tuber yield (Figure 1A) per plant than WT plants ( $p < 0.05$ ). The total biomass partitioned to the tubers (estimated as the percentage of total biomass (DW) partitioned in tubers) is consistently 15–25% higher in the *CDF1*-overexpressing plants than in the WT ones (Figure S2F), suggesting an increased partition of photoassimilates in tuber development in the transgenic plants.

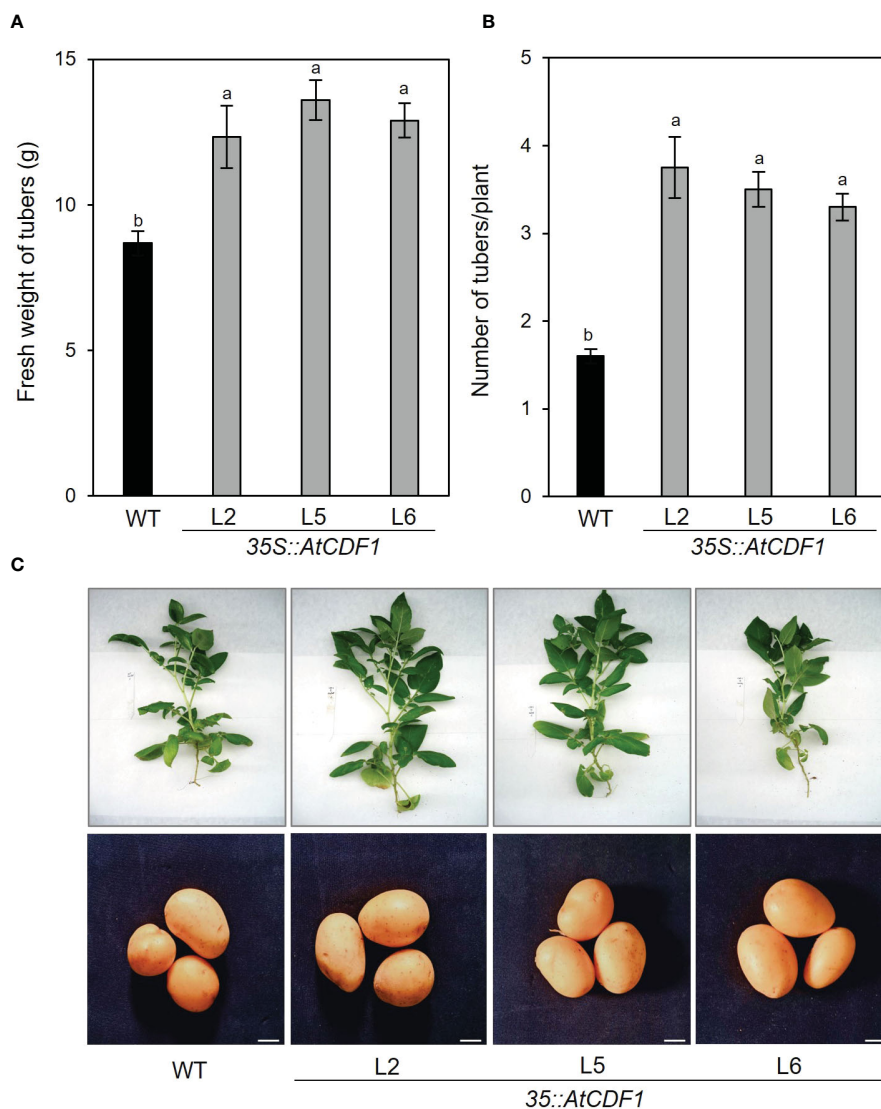


FIGURE 1

Tuber production of *35S::AtCDF1* and WT potato plants grown under growth chamber conditions. (A) Tuber yield (Fresh weight (FW) of tubers/plant) and (B) number of tubers per plant of overexpressing *35S::AtCDF1* (L2, L5 and L6 lines) and untransformed control (WT) potato (cv. Desiree) plants grown in growth chambers. (C) Representative pictures of plants and tubers of the analyzed lines. Data are expressed as means  $\pm$  SE of 8 to 10 plants. Different letters above the bars indicate a significant difference for a given genotype between WT and *35S::AtCDF1* plants ( $p < 0.01$ ) according to an analysis of variance, followed by a Student-Newman-Keuls test.

We also compared the agronomic performance of *35S::AtCDF1* (lines L5 and L6) and WT plants cultured under open field conditions by analysing both the total and the commercial (consumption and seed tubers) plant yield (Kg/ha) and the number of tubers per hectare (no tubers/ha). We found contrasting differences between the yield-related parameters of the transgenic and WT potato plants (Figure 2). As illustrated in Figure 2A, *35S::AtCDF1* plants produced 13-14% higher total and commercial-size tuber yields than the WT plants ( $p < 0.05$ ). In terms of productivity per land surface unit, control plants produced an average of 64 tonnes of tubers/hectare, whereas *AtCDF1*-overexpressing plants produced about 73 tonnes of tuber/hectare (Figure 2A).

Notably, although we did not observed any significant differences ( $p < 0.05$ ) in the number of tubers (n° tubers/ha)

between *35S::AtCDF1* and WT plants (Figure 2B), we found significant differences ( $p < 0.05$ ) in the size distribution of the tubers (Figure 2C). These were graded into three tuber size classes: out grade (0-25 mm), seed (25-40 mm) and consumption (greater than 40 mm). As shown in Figure 2C, the overexpressor lines produced 45-53% more tubers of the consumption size grade (Figure 2C) than the WT plants. However, it was also noticed that *35S::CDF1* lines showed a reduction of 8-15% and 37-51% in the number of seed and out grade tubers (smaller size categories) than those of the WT plants, respectively (Figure 2C). Moreover, the mean tuber weights of the *35S::CDF1* overexpressor plants (L5 and L6 lines), were ca. 12-23% and 17-19% heavier in the total and commercial categories than those of the WT, respectively ( $p < 0.05$ ; Figure 2D). Overall these data indicated that under field conditions the agronomic performance of *35S::AtCDF1* plants was better and

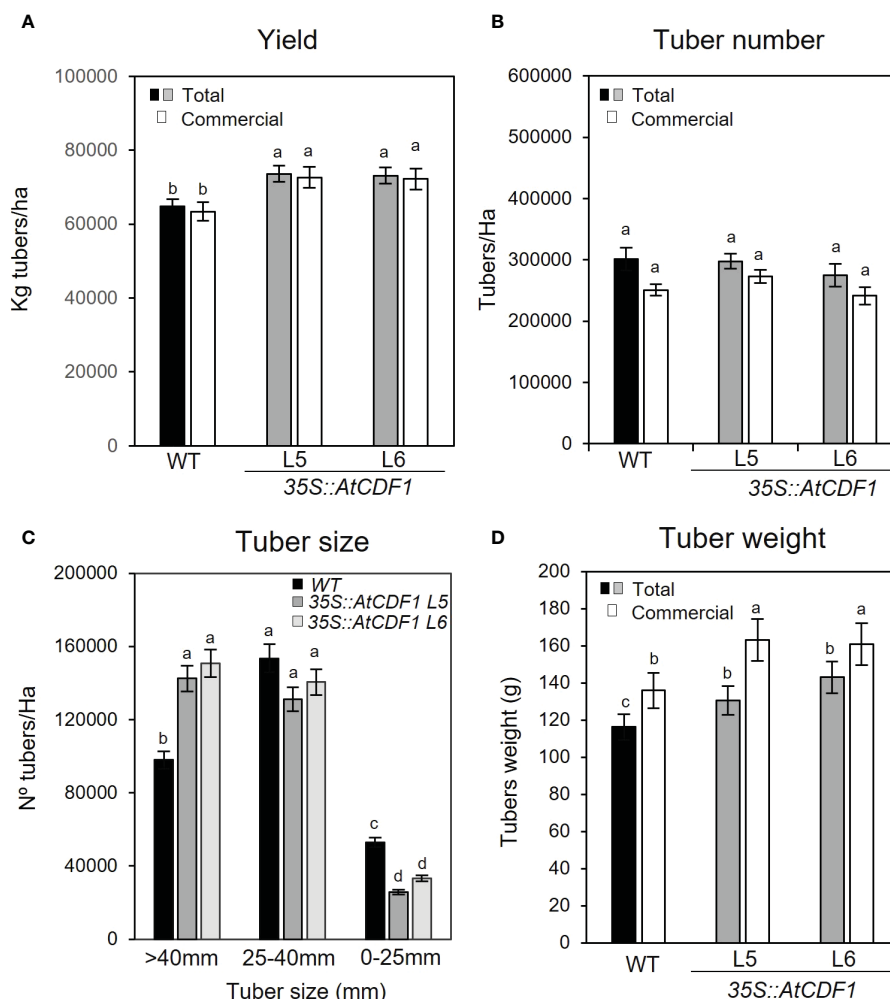


FIGURE 2

Agronomic performance of *35S::AtCDF1* and WT potato plants under field conditions. (A) Tuber yield (Kg tubers/ha) and (B) tuber number (no tubers/ha) of total and commercial tubers in untransformed control (WT; cv Desiree) and *35S::AtCDF1* (L5 and L6 lines) potato plants. (C) Number of tubers in 3 size categories: 0-25 mm (out grade), 25-40 mm (seed) and greater than 40 mm (consumption). (D) Mean tuber weight (g FW) of total and commercial tubers. Data are expressed as means  $\pm$  SE of 10 plants. Scale bar 0.5 cm. Significant differences between WT and *35S::AtCDF1* lines are indicated with different letters ( $p < 0.05$ ) according to an analysis of variance, followed by a Student-Newman-Keuls test.

demonstrated consistent improvement as compared to WT plants. In addition, the higher yield of *35S::AtCDF1* is likely due to an increased partition of biomass to the tubers, allowing the development of a higher number of bigger size tubers of the consumption size grade.

## The ectopic expression of *AtCDF1* promotes starch and amino acid accumulation in the tubers of plants cultured under open field conditions

Previously, we showed that the overexpression of *AtCDF3*, the closest *AtCDF1* homolog, in tomato, modified the amino acid and sugar contents of fruits (major sink organ) and resulted in enhanced yield and NUE under greenhouse conditions (Renau-Morata et al.,

2017; Domínguez-Figueroa et al., 2020). These results suggest that CDF3 plays an important role in both N and C assimilation and photoassimilates partition; and thus, CDFs might be used as a biotechnological target for the purposes of improving crop yields.

To examine the possible effects of *AtCDF1* ectopic expression on the C and N metabolism of potato tubers, as a major sink organ, we compared the contents of soluble sugars, starch and amino acids in tubers of *35S::AtCDF1* plants and WT plants cultured under open field conditions. As shown in Figure 3, *35S::AtCDF1* tubers accumulated similar levels of sucrose and fructose, but contained 30-39% higher levels of glucose than the WT tubers. Notably, the starch and amylose contents in *35S::AtCDF1* tubers were ca. 25% and 17% higher than in WT tubers, respectively. These data suggest that CDFs may be upstream regulators of starch biosynthesis pathways likely modulating, directly or indirectly, the activity of key C metabolic proteins. In addition, it has been shown that the

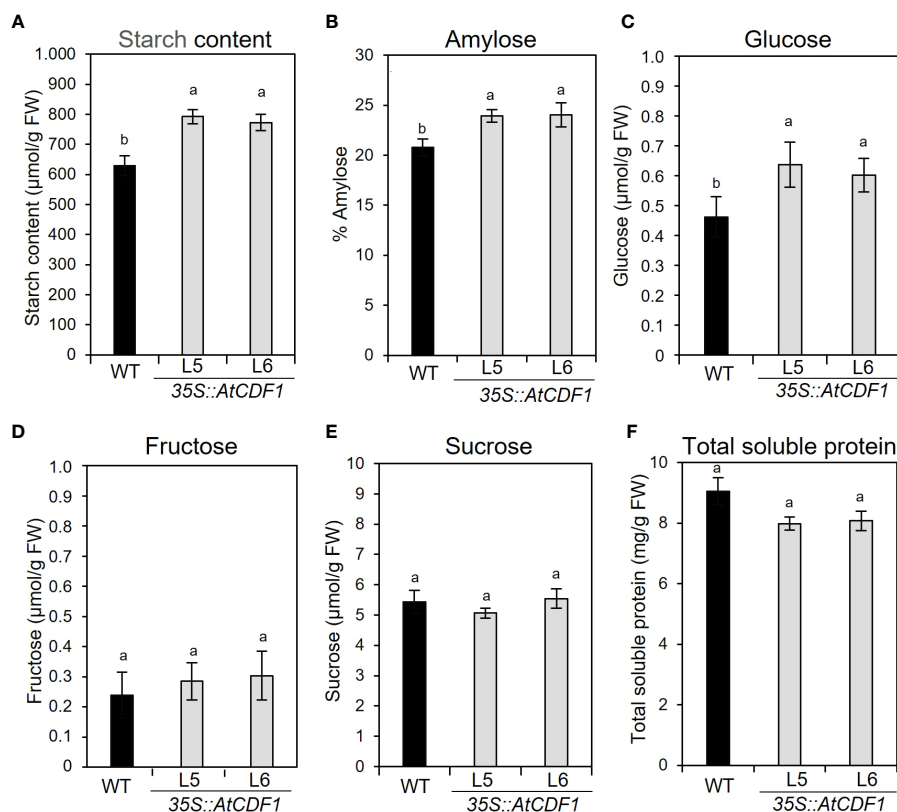


FIGURE 3

Starch, amylose, soluble sugars and protein contents in tubers of *35S::AtCDF1* and WT plants. (A) Starch content, (B) amylose percentage in starch, (C) glucose, (D) fructose and (E) sucrose contents, and (F) total soluble protein in tubers *35S::AtCDF1* (L5 and L6 lines) and WT (cv. Desiree) plants grown under field conditions. Data are expressed as means  $\pm$  SE of six replicates. Different letters above the bars indicate a significant differences between WT and *35S::AtCDF1* lines ( $p < 0.05$ ) according to an analysis of variance, followed by a StudentNewman-Keuls test.

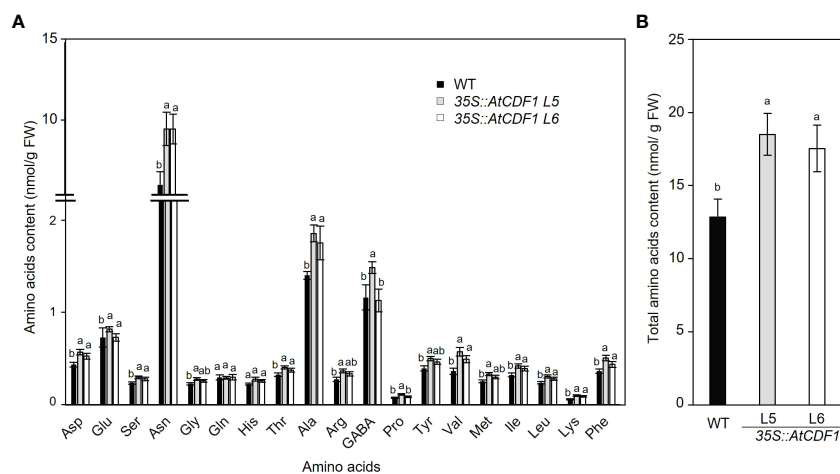
tuber can import but also conduct the “*de novo*” biosynthesis of all amino acids (Roessner et al., 2001; Roessner-Tunali et al., 2003). Therefore, we decided to investigate the impact of *AtCDF1* on the amino acid content in tubers of *AtCDF1* overexpressor and WT plants grown under field conditions. An analysis by GC-MS of the pool sizes of the different amino acids uncovered significant increases in most of the amino acids, especially asparagine, valine, isoleucine and alanine ( $p < 0.05$ ; Figure 4A). Consistently, the total amino acid content in the *35S::AtCDF1* tubers were ca. 22-58% higher than in the WT tubers (Figure 4B).

It is well known that the levels of adenylate pools are important factors that control/limit the starch content and yield and the concentration of different amino acids (Regierer et al., 2002; Carrari et al., 2005). In order to study the effects of the overexpression of *AtCDF1* on adenylate pools in tubers, we determined the steady-state levels of all three metabolites ATP, ADP, and AMP. The overexpression of *AtCDF1* led to significant changes in the levels of the various adenylate pools (Figure 5). Notably, the ATP and AMP contents in the *AtCDF1*-overexpressing tubers were ca. 4-5% and 12-16% higher than in ones in the WT tubers; but no significant changes were observed in the ADP levels

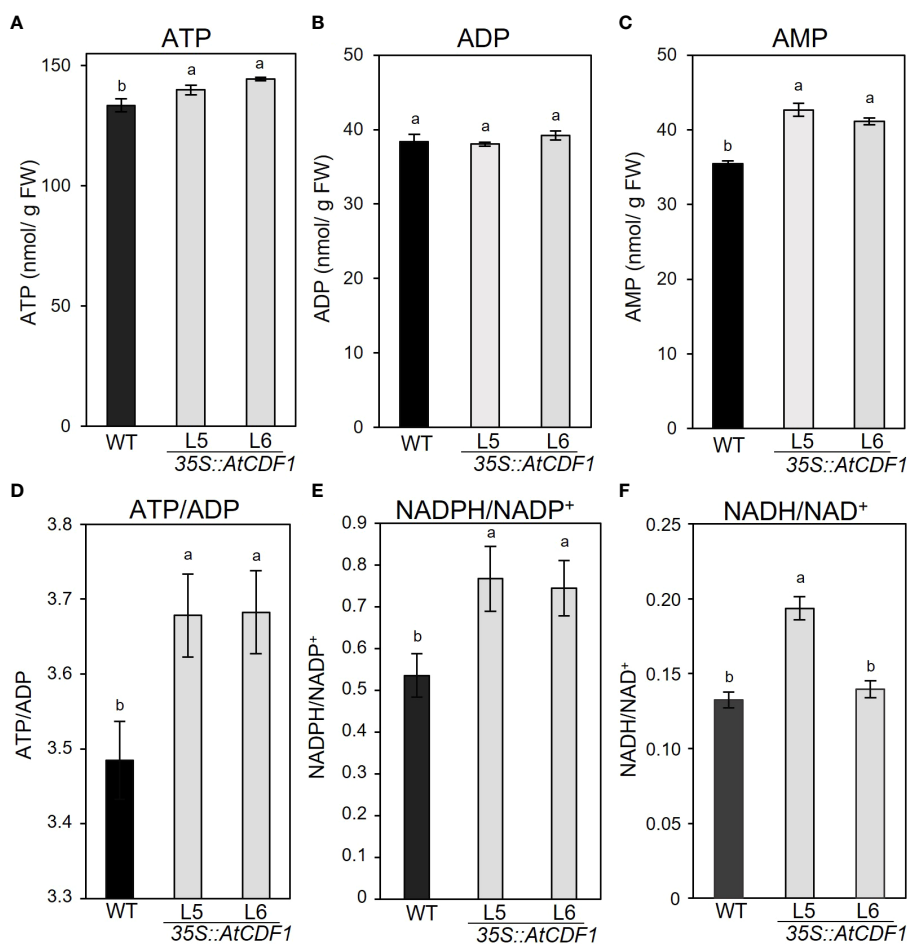
of the *35S::AtCDF1* lines (Figure 5). The ATP/ADP ratio is usually estimated as a central control parameter of cellular energy metabolism that determines the free-energy change for ATP hydrolysis and, therefore, the driving force for many reactions (Atkinson, 1968; Hardie and Hawley, 2001; Geigenberger et al., 2010). Thus we examined the ATP/ADP ratio and identified that were ca. 5-6% higher in tubers of *AtCDF1* overexpression plants than in the ones of the WT (Figure 5D), however, ATP/AMP ratio showed no significant changes (Figure S3).

We also analysed the content of the NAD(P)H/NAD(P) ratios. An accumulation of evidence has suggested that the nicotinamide adenine dinucleotide NADH/NAD<sup>+</sup> and NADPH/NADP<sup>+</sup> redox couples play crucial roles in the regulation of the cellular redox state, energy metabolism, mitochondrial function and gene expression, as well as in signalling pathways (Hashida and Kawai-Yamada, 2019; Smith et al., 2021); therefore this indicated that these redox couples are essential for the maintenance of a wide array of biological processes. Notably, NADPH/NADP<sup>+</sup> and NADH/NAD<sup>+</sup> ratios were ca. 39-43% and 1-29% higher in tubers of the *AtCDF1* overexpression plants (L5 and L6 lines) than in the WT (Figures 5E, F).





**FIGURE 4** Amino acids contents and composition in tubers of *35S::AtCDF1* and WT potato plants. (A) Amino acid composition profile and (B) total soluble amino acids in tubers of control (WT; cv Desiree) and *35S::AtCDF1* (L5 and L6 lines) potato plants grown under field conditions. Data are expressed as means  $\pm$  SE of nine independent replicates per line. For each amino acid, different letters above the bars indicate a significant difference to WT ( $p < 0.01$ ) according to an analysis of variance, followed by a Student-Newman-Keuls test.



**FIGURE 5** Levels of adenine nucleotides in potato tubers of WT and *35S::AtCDF1* plants. (A) ATP, (B) ADP, and (C) AMP contents, and (D) ATP/ADP, (E) NADPH/NADP<sup>+</sup> and (F) NADH/NAD<sup>+</sup> ratios of tubers of control (WT; cv. Desiree) and *35S::AtCDF1* plants (L5 and L6 lines) grown under field conditions. Data are expressed as means  $\pm$  SE of nine independent replicates per line. Significant differences between WT (cv Desiree) and transgenic plants are indicated with different letters ( $p < 0.05$ ) according to an analysis of variance, followed by a Student-Newman-Keuls test.

## The ectopic expression of *AtCDF1* in potato promotes significant changes in the tuber proteome

To further investigate the underlying mechanisms related to the observed effects of the ectopic expression of *AtCDF1* on tuber production and tuber quality parameters, we performed a differential proteomic analysis between tubers of *35S::AtCDF1* and WT plants grown under field conditions using LC-ESI MS/MS (deposited at ProteomeXchange Consortium PXD036489). These analyses revealed that the ectopic expression of *AtCDF1* promotes significant changes in the tuber proteome (Figure S4). As shown in Table S1, of the 407 differentially expressed proteins (DEPs) identified in this study ( $q$  value  $< 0.05$ ), 213 were up-regulated and 194 were down-regulated by the ectopic expression of *AtCDF1* (Figures 6, 7).

To identify differentially expressed pathways in *35S::AtCDF1* and WT tubers, a gene ontology (GO) term enrichment analysis according to molecular function (GO MF), biological process (GO BP) and cellular compartment (GO CC) was performed on the differentially expressed proteins. Substantially regulated GO terms are shown in Figures 6, 7. The list of all the GO terms associated with the regulated proteins is provided in Table S2. The over-representation analysis of biological functions associated with DEPs significantly up-regulated in *35S::AtCDF1* tubers revealed an enrichment in GO terms related to different aspects of primary metabolism including “ATP metabolic process”, “generation of precursors metabolites and energy”, “NADPH regeneration”, “carbohydrate derivative metabolic process”, “glucose-6-phosphate metabolic process” and “pentose-phosphate pathway” (Figure 6; Table S2). Specifically, we identified 81 proteins including

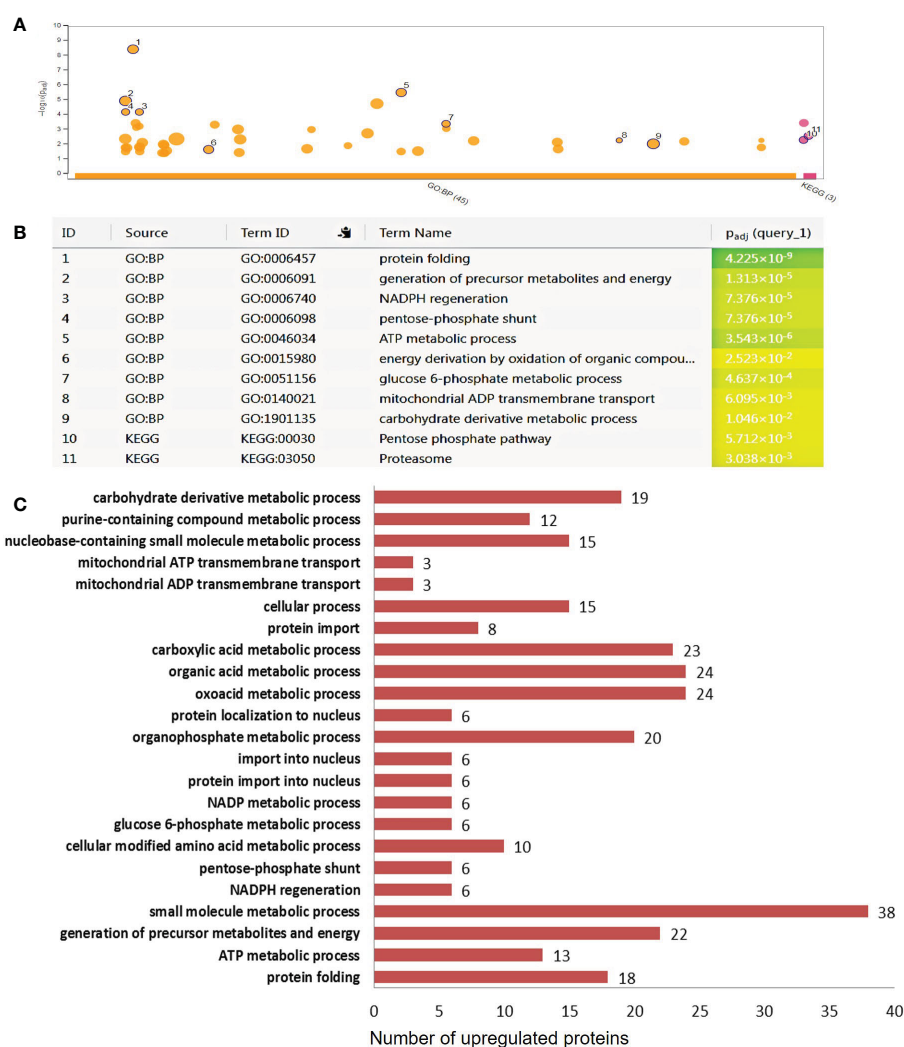


FIGURE 6

Functional categorization of the differentially upregulated proteins in *35S::AtCDF1* potato tubers. The significantly up regulated proteins as compared with control (WT, cv. Desiree) were organized according to their functional category using gPROFILER software (Reimand et al., 2016). (A) The results of enrichment analysis (GO BP and KEGG) were shown in a Manhattan plot. The x-axis indicates functional terms grouped by database used (color code) and the y-axis indicates the enrichment adjusted  $p$ -values. The dots shows all the enriched terms ( $p < 0.05$ ). The highlighted dots show GO terms emphasized in the text. The graphs (B, C) show the enriched terms highlighted in the Manhattan plot as well as the statistical significance ( $p$ -value) and the number of DEPs included in the enriched term (located close to the bar). The full details of the GO analysis are given in Supplementary Table S2.

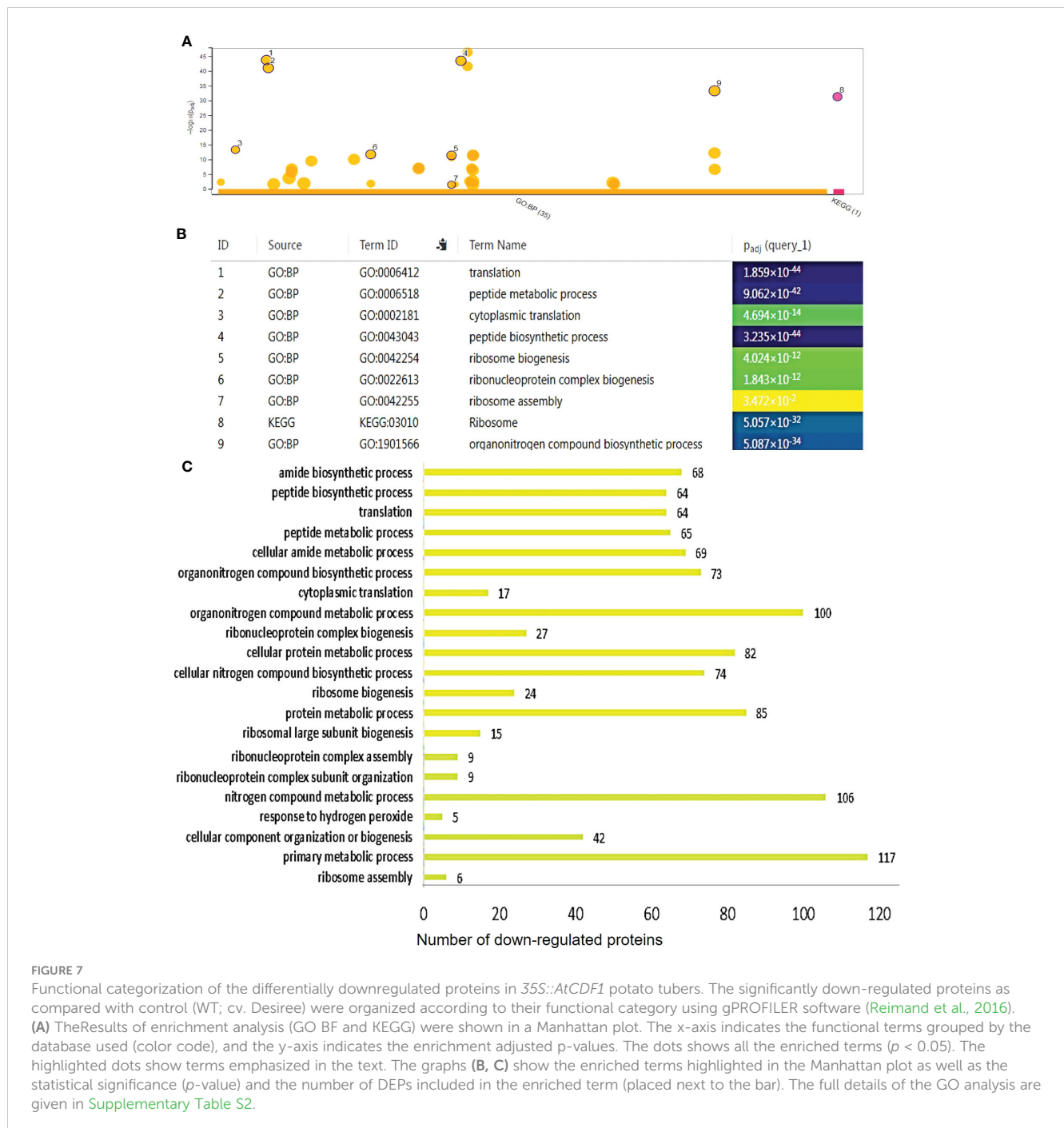


FIGURE 7

Functional categorization of the differentially downregulated proteins in *35S::AtCDF1* potato tubers. The significantly down-regulated proteins as compared with control (WT; cv. Désirée) were organized according to their functional category using gPROFILER software (Reimand et al., 2016). (A) The results of enrichment analysis (GO BF and KEGG) were shown in a Manhattan plot. The x-axis indicates the functional terms grouped by the database used (color code), and the y-axis indicates the enrichment adjusted p-values. The dots show all the enriched terms ( $p < 0.05$ ). The highlighted dots show terms emphasized in the text. The graphs (B, C) show the enriched terms highlighted in the Manhattan plot as well as the statistical significance ( $p$ -value) and the number of DEPs included in the enriched term (placed next to the bar). The full details of the GO analysis are given in Supplementary Table S2.

ADP/ATP mitochondrial carrier proteins, ATP synthase subunits and the enzymes involved in sucrose/starch metabolism and pentose phosphate pathway (e.g. sucrose synthase [SuSy], granule-bound starch synthase [GBSS] 1, SNF1-related protein kinase regulatory subunit gamma-1 [SnRK1], transaldolase and 6-phosphogluconolactonase) (Table S1). These analyses also identified the different up-regulated DEPs involved in various aspects of oxidative metabolism, including glycolysis (e.g. pyrophosphate fructose-6-phosphate-1-phosphotransferase, fructose-bisphosphate aldolase, pyruvate kinase and enolase) and TCA cycle (e.g. citrate synthase and malic enzyme), as well as PEP carboxylase (Table S1). Moreover, we found GO terms related to

nitrogen metabolism and protein folding that are known to be regulated by CDFs (Renau-Morata et al., 2020a). Specifically, we found an enrichment in terms such as “cellular modified amino acid metabolic process” and “protein folding”, including proteins like glutamate decarboxylase (involved in GABA biosynthesis) and betaine aldehyde dehydrogenase, and Heat shock 70, 80 proteins and chaperonins (Figure 6; Tables S1, S2), as well as a group of storage proteins including several patatin proteins (Table S1). The GO term analysis of the down-regulated proteins revealed the overrepresentation of terms associated with translation, ribosome biogenesis and protein biosynthesis such as “ribosome assembly”, “ribosome biogenesis”, “translation”, “peptide biosynthetic process”

and “ribosome” (Table S2). Specifically, we identified a group of downregulated translation- and ribosome biogenesis-related proteins including elongation factors-1- $\alpha$ , -3, eukaryotic translation initiation factor 5A, lysyl/seryl-tRNA synthetases and ribosomal proteins such as L9-1, L13, L26, L27a, S30 and S16 (Figure 7; Table S1). This analysis also revealed that 35S::*AtCDF1* tubers accumulated lower levels of the enzymes involved in starch breakdown (e.g. starch phosphorylase and alpha-amylase) than WT tubers (Table S1).

## Discussion

DOF TFs have been extensively explored in various model plants and crops (Yanagisawa, 2016; Renau-Morata et al., 2020a). Of these, CDF TFs are known to be involved in the control of different facets of plant growth and responses to abiotic stresses (Corrales et al., 2014; Fornara et al., 2015; Corrales et al., 2017). In this study, we assessed the impact of the ectopic expression of the Arabidopsis *CDF1* gene on tuber composition and the yield and growth of potato plants cultured under both growth chamber and open field conditions.

### The ectopic expression of *AtCDF1* enhances the tuber yield and quality in potato plants by increasing tuber sink strength

The ectopic expression of *AtCDF1* in potato did not alter the photosynthetic capacity or the total biomass of the plant, but significantly enhanced tuber yield when the plants were cultured in growth chambers (Figures 1, S1, S2). This clearly indicates that CDFs could play an important role in the control of photoassimilates carbon partitioning by increasing tuber sink strength. The data obtained in this study are coherent with previous results pertaining to tomato and Arabidopsis CDFs, since the ectopic expression of *AtCDF3* and *SICDF3* in tomato promoted higher yield and improved the fruit's amino acid and sugar contents (Renau-Morata et al., 2017). Moreover, the targeted overexpression of the tomato *SICDF4* gene in the fruit increased the fruit size and yield, which were related to a greater amount of both water and dry matter accumulated in the fruit (Renau-Morata et al., 2020b).

Under both growth chamber and field conditions, 35S::*AtCDF1* lines exhibited significant increases in tuber yield (Figures 1, 2). However, our study revealed differences in the tuber traits depending on the experimental conditions. The higher yield of the overexpressor lines in the growth chambers was related to a higher number of developed tubers. However, the number of tubers in the field were similar in both the WT and 35S::*CDF1* plants and the greater yield was due to an increase in both the mean size and weight of the tubers. Similarly, other analyses of potato indicated

that the cultivation conditions clearly influence growth-related traits. The tuber yield and the aboveground biomass differ depending on whether they are greenhouse or field conditions (Pourazari et al., 2018). Therefore, due to the marked effect of the environment on targeted and non-targeted traits in GM plants, the performance of the lines has to be verified under commercial management conditions (Glandorf and Breyer, 2016). In line with this presumption, different molecular analyses have demonstrated that *CDF* gene expression is differentially modulated by the environmental conditions (Imaizumi et al., 2005; Sawa et al., 2007; Kloosterman et al., 2013; Corrales et al., 2017; Renau-Morata et al., 2020a).

Notably, here we found that the ectopic expression of *AtCDF1* triggers significant changes in carbon metabolism (Figures 3, 6, 7). Tubers of plants overexpressing *AtCDF1* accumulated ca. 25% more glucose and starch, thus improving the nutritional value of the tuber for food. In addition, a higher amylose content (ca. 17% compared to WT) was also found in the transgenic tubers (Figure 3). Amylose content is a valuable trait associated with different industrial applications of starch. In fact, “high amylose” starches tend to have greater retrogradation rates following processing than those that have a high amylopectin content (Singh and Anderson, 2004). In addition, high amylose starches show high freeze-thaw quality (Jobling et al., 2002) and reduced oil penetration, which are highly valuable tools in the food processing industry (Tarn et al., 2006). Consequently, developing potato cultivars with a high amylose content is an active research area in crop biotechnology (Santelia and Zeeman, 2010; Patil et al., 2016; Dahal et al., 2019);.

The amino acid content is also an important determinant of the tuber's nutritional and processing quality (Brierley et al., 1996). The ectopic expression of *AtCDF1* promoted significant increases in the contents of essential amino acids (Reeds, 2000), including valine, phenylalanine, threonine, methionine, leucine, isoleucine and lysine in tubers. It is interesting to point out that valine, methionine and leucine are limiting amino acids in the commercial isolated potato protein (Jiménez-Munoz et al., 2022 and references therein). Besides, 35S::*AtCDF1* tubers exhibited higher levels of expression of a group of proteins involved in proteolytic pathways (Figure 6; Table S1). It is, thus, conceivable that *CDF1* mediates the tuber's amino acid content and nutritional quality through the action on protein turnover and degradation-related pathways. In addition, it is also possible that *AtCDF1* might play a role in the control of amino acid translocation from source to sink organs. Interestingly, the 35S::*AtCDF1* tubers showed higher levels of patatin proteins than the WT ones (Table S1), which are recognized as important nutritional components of potato tubers (Bárta and Bártová, 2009), and have properties such as foaming, emulsifying and enzymatic activities, which are of interest in a number of different biotechnological applications (Ralet and Gueguen, 2001; Van Koningsveld et al., 2006). Overall, the data presented in this study show that the ectopic expression of *AtCDF1* can be used to improve the yield and quality properties and breeding of potato for different industrial applications.

## Overexpression of *AtCDF1* gene results in major changes in the tuber proteome

Up to now, previous studies have mainly used transcriptomic approaches to investigate the molecular mechanism of action of CDFs (Fornara et al., 2009; Yanagisawa, 2016; Corrales et al., 2017; Renau-Morata et al., 2017; Renau-Morata et al., 2020a). However, transcript abundance on its own cannot be utilized to directly deduce directly changes in the proteome and/or metabolic activity/fluxes in the plant metabolism (Nakaminami et al., 2014; Schwender et al., 2014). In this study, we conducted comparative proteomic analysis between 35S::*AtCDF1* and WT tubers and identified a group of 407 differentially expressed proteins (DEPs) involved in different aspects of C and N metabolism (Figures 6, 7; S6; Tables S1, S2). Consistent with the above data showing that the *AtCDF1* ectopic expression enhances the potato starch content and yield, these analyses revealed that the ectopic expression of *AtCDF1* upregulated the expression of strong determinants of sink strength, as well as tuber starch content, tuber quality and yield, such as SuSy and GBSS 1 and 2 (Hovenkamp-Hermelink et al., 1987; Baroja-Fernández et al., 2009; Li et al., 2013; Toinga-Villafuerte et al., 2022). In addition, the expression of enzymes involved in starch breakdown, such as starch phosphorylase and alpha-amylase, were downregulated (Tables S1, S2). This indicates that CDFs act as upstream regulators of C metabolism. In line with this hypothesis, previous reports have shown that the ectopic expressions of Arabidopsis *AtCDF3* and tomato *SICDF3* in tomato impacts C/N metabolism genes, promoting a rise in sugar and amino acids contents, thus allowing increased growth and yield (Renau-Morata et al., 2017; Domínguez-Figueroa et al., 2020). Moreover, the targeted overexpression of the tomato *SICDF4* gene in the fruit led to a larger amount of biomass been partitioned to the fruit; this was based on the greater sink strength of the fruits induced by the increased activity of enzymes involved in the sucrose and starch metabolism, including invertase, SuSy, ADP-glucose pyrophosphorylase (AGP) and UDP-glucose pyrophosphorylase (Renau-Morata et al., 2020b).

The proteomic analyses also revealed that the ectopic expression of *AtCDF1* upregulated the expression of SnRK1 and 14-3-3 proteins. The SnRK1 plays an important role in the regulation of development and metabolism in potato tubers and thus yield and quality (McKibbin et al., 2006; Crozet et al., 2014; Wurzinger et al., 2018). 14-3-3 proteins regulate key process in C/N metabolism by direct binding and fine-tuning the activity of a wide array of essential enzymes involved in carbohydrate and nitrogen metabolism, such as nitrate reductase, sucrose phosphate synthase, AGP, glutamine synthetase or H<sup>+</sup>-ATPase (Oecking et al., 1994; Bachmann et al., 1996a; Toroser et al., 1998; Moorhead et al., 1999; Sehne et al., 2001; Comparot et al., 2003; Chevalier et al., 2009). In addition, the overexpression of different 14-3-3 genes in potato disturbed the sugar, catecholamine and lipid contents. In contrast, a 14-3-3 antisense affected the tuber starch content, nitrate reductase activity and amino acid composition (Prescha et al., 2001; Swiedrych et al., 2002; Zuk et al., 2005). Overall, our data suggest

that the ectopic expression of *AtCDF1* affects primary metabolism, at least in part, through SnRK1 and 14-3-3 regulatory proteins.

It is well known that the adenylate pools determine the starch content and yield and the accumulation of different amino acids (Tjaden et al., 1998; Loef et al., 2001; Regierer et al., 2002; Carrari et al., 2005). In plant heterotrophic tissues, the energy metabolism and adenylate energy charge are determined by cell respiration pathways (van Dongen et al., 2011). The proteomic analyses revealed that the ectopic expression of *AtCDF1* upregulates the glycolytic and pentose phosphate pathways, the tricarboxylic acid (TCA) cycle and the mitochondrial electron transport chain, as well as adenylate kinases and the ADP-ATP mitochondrial carrier (Tables S1, S2; Figure S5). These data would support the higher ATP content and NAD(P)H/NAD(P)<sup>+</sup> ratios in 35S::*AtCDF1* tubers than in the WT ones (Figure 5) which could be channelled for the biosynthesis of starch and amino acids in the tubers.

Overall, these data might suggest that CDF1 may regulate the expression of genes encoding the key enzymes involved in primary metabolism, in all likelihood by direct binding to their promoter regions. The examination of the promoter regions of the highlighted DEPs involved in different processes, such as the ATP metabolism, the enzymes involved in sucrose/starch metabolism, the pentose phosphate pathway, the TCA cycle, as well as the genes involved in amino acid metabolism, ribosome biogenesis and protein folding, consistently showed multiple putative DOF binding sites in their promoter regions (Figure S6), suggesting that these groups of genes might be direct targets of CDF1.

In conclusion, the present study showed that the expression of the *CDF1* gene from Arabidopsis increased the yield of potato plants under both chamber and field conditions, and enhanced the tuber quality properties for different industrial applications. The increased yield observed was associated with changes in photoassimilate partitioning and C and N metabolism, as well as with changes in the expression of the proteins involved in energy production and sucrose and starch metabolisms. Together with what was previously described for tomatoes (Renau-Morata et al., 2017; Domínguez-Figueroa et al., 2020; Renau-Morata et al., 2020b), these data highlight the CDFs as potential breeding tools for improving the sink strength of the harvested potato organs in the *Solanaceae*. Furthermore, the higher yields promoted by CDFs were obtained under the highly heterogeneous field conditions.

## Data availability statement

The data presented in the study are deposited in the ProteomeXchange repository, accession number PXD036489.

## Author contributions

EB-F, JV-C, SGN, RVM and JM designed, planned, and organized the experiments. LC, EB-F, BR-M, FJM, JC, SC, LY, AS-L and MGC

performed the research. SGN, RVM, JP-R and JM wrote the article. All authors contributed to the article and approved the submitted version.

## Acknowledgments

We thank Manuel Sánchez-Perales and Antonio Ramos for his technical assistance and Mike Bennet for English editing. This study was supported by the Ministerio de Ciencia e Innovación (MCIN) and the Agencia Estatal de Investigación (AEI) / 10.13039/501100011033/ (grants RTA2015-00014-c02-00 to JM and SGN; PID2020-114165RR-C21 to JM), (BIO2017-82873-R to JVC) (PID2019-104685GB-I00 to JP-R, EB-F and FJM), the Gobierno de Navarra (PC036-037 BIOMEF to EB-F and FJM) and the Vicerrectorado de Investigación de la Universitat Politècnica de València (PAID-11-21 to SGN), J.C is funded by the National Agency for Research and Development (ANID) Chile with Program FONDECYT Regular 1190812 and ANID-Millennium Science Initiative Program-ICN17\_022" and The GEP Chinese pre-doctoral fellowship to LY. We also want to acknowledge the "Severo Ochoa Program for Centres of Excellence in R&D" (CEX2020-000999-S) supported by MCIN/AEI/10.13039/501100011033 for supporting the scientific services used in this study.

## References

- Abelenda, J. A., Bergonzi, S., Oortwijn, M., Sonnewald, S., Du, M., Visser, R. G. F., et al. (2019). Source-sink regulation is mediated by interaction of an FT homolog with a SWEET protein in potato. *Curr. Biol.* 29 (7), 1178–1186. doi: 10.1016/j.cub.2019.02.018
- Alvarez, J. M., Schinke, A. L., Brooks, M. D., Pasquino, A., Leonelli, L., Varala, K., et al. (2020). Transient genome-wide interactions of the master transcription factor NLP7 initiate a rapid nitrogen-response cascade. *Nat. Commun.* 11, 1157. doi: 10.1038/s41467-020-14979-6
- Andersson, M., Melander, M., Pojmark, P., Larsson, H., Bulow, L., and Hofvander, P. (2006). Targeted gene suppression by RNA interference: An efficient method for production of high-amylose potato lines. *Journal of Biotechnology*, 123:137–148.
- Atkinson, D. E. (1968). Energy charge of the adenylate pool as a regulatory parameter. interaction with feedback modifiers. *Biochem.* 7 (11), 4030–4034. doi: 10.1021/bi00851a033
- Bachmann, M., Huber, J. L., Athwal, G. S., Wu, K., Ferl, R. J., and Huber, S. C. (1996). 14-3-3 proteins associate with the regulatory phosphorylation site of spinach leaf nitrate reductase in an isoform-specific manner and reduce dephosphorylation of ser-543 by endogenous protein phosphatases. *FEBS Letts.* 398, 26–30. doi: 10.1016/s0014-5793(96)01188-x
- Bahaji, A., Baroja-Fernández, E., Ricarte-Bermejo, A., Sánchez-López, Á.M., Muñoz, F. J., Romero, J. M., et al. (2015). Characterization of multiple SPS knockout mutants reveals redundant functions of the four arabidopsis sucrose phosphate synthase isoforms in plant viability, and strongly indicates that enhanced respiration and accelerated starch turnover can alleviate the blockage of sucrose biosynthesis. *Plant Sci.* 238, 135–147. doi: 10.1016/j.plantsci.2015.06.009
- Baroja-Fernández, E., Muñoz, F. J., Montero, M., Etxebarria, E., Sesma, M. T., Ovecka, M., et al. (2009). Enhancing sucrose synthase activity in transgenic potato (*Solanum tuberosum* L.) tubers results in increased levels of starch, ADPglucose and UDPglucose and total yield. *Plant Cell Physiol.* 50 (9), 1651–1662. doi: 10.1093/pcp/pcp108
- Baroja-Fernández, E., Muñoz, F. J., Saikusa, T., Rodríguez-López, M., Akazawa, T., and Pozueta-Romero, J. (2003). Sucrose synthase catalyzes the *de novo* production of ADPglucose linked to starch biosynthesis in heterotrophic tissues of plants. *Plant Cell Physiol.* 44 (5), 500–509. doi: 10.1093/pcp/pcg062
- Barrell, P. J., Meiyalaghan, S., Jacobs, J. M., and Conner, A. J. (2013). Applications of biotechnology and genomics in potato improvement. *Plant Biotech. J.* 11 (8), 907–920. doi: 10.1111/pbi.12099
- Bárta, J., and Bártoová, V. (2009). Patatin, the major protein of potato (*Solanum tuberosum* L.) tubers, and its occurrence as genotype effect: Processing versus table potatoes. *Czech J. Food Sci.* 26, 347–359. doi: 10.17221/27/2008-CJFS
- Birch, P. R. J., Bryan, G., Fenton, B., Gilroy, E. M., Hein, I., Jones, J. T., et al. (2012). Crops that feed the world 8: Potato: are the trends of increased global production sustainable? *Food Secur.* 4, 477–508. doi: 10.1007/s12571-012-0220-1
- Bradshaw, J. E. (2021). *Potato breeding: theory and practice* (Switzerland: Springer). doi: 10.1007/978-3-030-64414-7
- Brierley, E. R., Bonner, P. L. R., and Coob, A. H. (1996). Factors influencing the free amino acid content of potato (*Solanum tuberosum* L.) tubers during prolonged storage. *J. Sci. Food Agric.* 70, 515–525. doi: 10.1002/(SICI)1097-0010
- Bulley, S., Wright, M., Rommens, C., Yan, H., Rassam, M., Lin-Wang, K., et al. (2012). Enhancing ascorbate in fruits and tubers through over-expression of the l-galactose pathway gene GDP-l-galactose phosphorylase. *Plant Biotech. J.* 10 (4), 390–397. doi: 10.1111/j.1467-7652.2011.00668.x
- Burgos, G., Zum Felde, T., Andre, C., and Kubow, S. (2020). "The potato and its contribution to the human diet and health," in *The potato crop*. Eds. H. Campos and O. Ortiz (Switzerland: Springer), 37–74. doi: 10.1007/978-3-030-28683-5
- Carrari, F., Coll-Garcia, D., Schauer, N., Lytovchenko, A., Palacios-Rojas, N., Balbo, I., et al. (2005). Deficiency of a plastidial adenylate kinase in arabidopsis results in elevated photosynthetic amino acid biosynthesis and enhanced growth. *Plant Physiol.* 137 (1), 70–82. doi: 10.1104/pp.104.056143
- Chakraborty, S., Chakraborty, N., Agrawal, L., Ghosh, S., Narula, K., Shekhar, S., et al. (2010). Next-generation protein-rich potato expressing the seed protein gene AmA1 is a result of proteome rebalancing in transgenic tuber. *Proc. Nat. Acad. Sci. U.S.A.* 107 (41), 17533–17538. doi: 10.1073/pnas.1006265107
- Chevalier, D., Morris, E. R., and Walker, J. C. (2009). 14-3-3 and FHA domains mediate phosphoprotein interactions. *Annu. Rev. Plant Biol.* 60, 67–91. doi: 10.1146/annurev-arplant.59.032607.092844
- Comparot, S., Lingiah, G., and Martin, T. (2003). Function and specificity of 14-3-3 proteins in the regulation of carbohydrate and nitrogen metabolism. *J. Exp. Bot.* 54, 595–604. doi: 10.1093/jxb/erg057
- Corrales, A. R., Carrillo, L., Lasierra, P., Nebauer, S. G., Domínguez-Figueroa, J., Renau-Morata, B., et al. (2017). Multifaceted role of cycling dof factor 3 (CDF3) in the regulation of flowering time and abiotic stress responses in arabidopsis. *Plant Cell Environ.* 40, 748–764. doi: 10.1111/pce.12894
- Corrales, A. R., Nebauer, S. G., Carrillo, L., Fernández-Nohales, P., Marqués, J., Renau-Morata, B., et al. (2014). Characterization of tomato cycling dof factors reveals conserved and new functions in the control of flowering time and abiotic stress responses. *J. Exp. Bot.* 65, 995–1012. doi: 10.1093/jxb/ert451
- Crozet, P., Margalha, L., Confraria, A., Rodrigues, A., Martinho, C., Adamo, M., et al. (2014). Mechanisms of regulation of SNF1/AMPK/SnRK1 protein kinases. *Front. Plant Sci.* 5. doi: 10.3389/fpls.2014.00190

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2023.1010669/full#supplementary-material>

- Dahal, K., Li, X.-Q., Tai, H., Creelman, A., and Bizimungu, B. (2019). Improving potato stress tolerance and tuber yield under a climate change scenario—a current overview. *Front. Plant Sci.* 10. doi: 10.3389/fpls.2019.00563
- Dancs, G., Kondrák, M., and Bánfalvi, Z. (2008). The effects of enhanced methionine synthesis on amino acid and anthocyanin content of potato tubers. *BMC Plant Biol.* 8 (1), 1–10. doi: 10.1186/1471-2229-8-65
- Domínguez-Figueroa, J., Carrillo, L., Renau-Morata, B., Yang, L., Molina, R.-V., Marino, D., et al. (2020). The arabidopsis transcription factor CDF3 is involved in nitrogen responses and improves nitrogen use efficiency in tomato. *Front. Plant Sci.* 11. doi: 10.3389/fpls.2020.601558
- FAO (2020) *FAOSTAT production database. latest update: 02/17/2022*. Available at: [www.fao.org/faostat](http://www.fao.org/faostat).
- Fornara, F., de Montaigu, A., Sánchez-Villarreal, A., Takahashi, Y., van Themaat, E. V. L., Huettel, B., et al. (2015). The GI-CDF module of arabidopsis affects freezing tolerance and growth as well as flowering. *Plant J.* 81 (5), 695–706. doi: 10.1111/tj.12759
- Fornara, F., Panigrahi, K. C., Gissot, L., Sauerbrunn, N., Rühl, M., Jarillo, J. A., et al. (2009). Arabidopsis DOF transcription factors act redundantly to reduce *CONSTANS* expression and are essential for a photoperiodic flowering response. *Dev. Cell.* 17 (1), 75–86. doi: 10.1016/j.devcel.2009.06.015
- Geigenberger, P., Riewe, D., and Fernie, A. R. (2010). The central regulation of plant physiology by adenylates. *Trends Plant Sci.* 15, 98–105. doi: 10.1016/j.tplants.2009.11.004
- Glandorf, D. C. M., and Breyer, D. (2016). “Field trials with GM trees: A step-by-step approach,” in *Biosafety of forest transgenic trees. forestry sciences*, vol. 82. (Dordrecht: Springer). doi: 10.1007/978-94-017-7531-1\_7
- Halterman, D., Guenther, J., Collinge, S., Butler, N., and Douches, D. (2016). Biotech potatoes in the 21st century: 20 years since the first biotech potato. *Am. J. Potato Res.* 93, 1–20. doi: 10.1007/s12230-015-9485-1
- Hameed, A., Zaidi, S. S., Shakir, S., and Mansoor, S. (2018). Applications of new breeding technologies for potato improvement. *Front. Plant Sci.* 9. doi: 10.3389/fpls.2018.00925
- Hardie, D. G., and Hawley, S. A. (2001). AMP-activated protein kinase: the energy charge hypothesis revisited. *Bioessays* 23, 1112–1119. doi: 10.1002/bies.10009
- Hashida, S. N., and Kawai-Yamada, M. (2019). Inter-organelle NAD metabolism underpinning light responsive NADP dynamics in plants. *Front. Plant Sci.* 26 (10). doi: 10.3389/fpls.2019.00960
- Haverkort, A. J., and Struik, P. C. (2015). Yield levels of potato crops: recent achievements and future prospects. *Field Crops Res.* 182, 76–85. doi: 10.1016/j.fcr.2015.06.002
- Hoagland, D. R., and Arnon, D. I. (1950). *The water-culture method for growing plants without soil* Vol. 347 (Berkeley, Calif: University of California, College of Agriculture, Agricultural Experiment Station).
- Hofvander, P., Ischebeck, T., Turesson, H., Kushwaha, S. K., Feussner, I., Carlsson, A. S., et al. (2016). Potato tuber expression of arabidopsis WRINKLED1 increase triacylglycerol and membrane lipids while affecting central carbohydrate metabolism. *Plant Biotech. J.* 14 (9), 1883–1898. doi: 10.1111/pbi.12550
- Hovenkamp-Hermelink, J. H. M., Jacobsen, E., Ponstein, A. S., Visser, R. G. F., Vos-Scheperkeuter, G. H., Bijmolt, E. W., et al. (1987). Isolation of an amylose-free starch mutant of the potato (*Solanum tuberosum* L.). *Theor. Appl. Genet.* 75, 217–221. doi: 10.1007/BF00249167
- Imaizumi, T., Schultz, T. F., Harmon, F. G., Ho, L. A., and Kay, S. A. (2005). FKF1 f-box protein mediates cyclic degradation of a repressor of *CONSTANS* in arabidopsis. *Science* 309, 293–297. doi: 10.1126/science.1110586
- Itkin, M., Heinig, U., Tzfadia, O., Bhidi, A. J., Shinde, B., Cardenas, P. D., et al. (2013). Biosynthesis of antinutritional alkaloids in solanaceous crops is mediated by clustered genes. *Science* 341, 175–179. doi: 10.1126/science.124023
- Jiménez-Munoz, L., Tsochatzis, E. D., and Corredig, M. (2022). Impact of the structural modifications of potato protein in the digestibility process under semi-dynamic simulated human gastrointestinal *In vitro* system. *Nutrients* 14, 2505. doi: 10.3390/nu14122505
- Jobling, S., Westcott, R., Tayal, A., Jeffcoat, R., and Schwall, P. (2002). Production of a freeze-thaw-stable potato starch by antisense inhibition of three starch synthase genes. *Nat. Biotech.* 20, 295–299. doi: 10.1038/nbt0302-295
- Kiel, M., Sanchez-Serrano, J. J., and Willmitzer, L. (1989). Both wound inducible and tuber specific expression mediated by the promoter of a single member of the protease inhibitor 11 gene families. *EMBO J.* 8, 1323–1330. doi: 10.1002/j.1460-2075.1989.tb03512.x
- Kloosterman, B., Abelenda, J. A., Mdel, M., Oortwijn, M., de Boer, J. M., Kowitwanich, K., et al. (2013). Naturally occurring allele diversity allows potato cultivation in northern latitudes. *Nature* 495 (7440), 246–250. doi: 10.1038/nature11912
- Kronic, S. L., Skryhan, K., Mikkelsen, L., Ruzanski, C., Shaik, S. S., Kirk, H. G., et al. (2018). Non-GMO potato lines with an altered starch biosynthesis pathway confer increased-amylose and resistant starch properties. *Starch* 70 (1–2), 1600310. doi: 10.1002/star.201600310
- Kumar, P., and Jander, G. (2017). Concurrent overexpression of arabidopsis thaliana cystathionine  $\gamma$ -synthase and silencing of endogenous methionine  $\gamma$ -lyase enhance tuber methionine content in *Solanum tuberosum*. *J. Agric Food Chem.* 65 (13), 2737–2742. doi: 10.1021/acs.jafc.7b00272
- Kurai, T., Wakayama, M., Abiko, T., Yanagisawa, S., Aoki, N., and Ohsugi, R. (2011). Introduction of the ZmDof1 gene into rice enhances carbon and nitrogen assimilation under low-nitrogen conditions. *Plant Biotechnol. J.* 9, 826–837. doi: 10.1111/j.1467-7652.2011.00592.x
- Li, J., Baroja-Fernández, E., Bahaji, A., Muñoz, F. J., Ovecka, M., Montero, M., et al. (2013). Enhancing sucrose synthase activity results in increased levels of starch and ADP-glucose in maize (*Zea mays* L.) seed endosperms. *Plant Cell Physiol.* 54 (2), 282–294. doi: 10.1093/pcp/pcs180
- Li, L., Yang, Y., Xu, Q., Owsiany, K., Welsch, R., Chitchumroonchokchai, C., et al. (2012). The or gene enhances carotenoid accumulation and stability during post-harvest storage of potato tubers. *Mol. Plant* 5 (2), 339–352. doi: 10.1093/mp/ssr099
- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2<sup>-ΔΔCT</sup> method. *Methods* 25, 402–408. doi: 10.1006/meth.2001.1262
- Loef, I., Stitt, M., and Geigenberger, P. (2001). Increased adenine nucleotide levels modify the interaction between respiration and starch synthesis when adenine is fed to tubers of growing potato tubers. *Planta* 212, 782–791. doi: 10.1007/s004250000461
- Loiret, F. G., Grimm, B., Hajirezaei, M. R., Kleiner, D., and Ortega, E. (2009). Inoculation of sugarcane with *Pantoea* sp. increases amino acid contents in shoot tissues: serine, alanine, glutamine and asparagine permit concomitantly ammonium excretion and nitrogenase activity of the bacterium. *J. Plant Physiol.* 166, 1152–1161. doi: 10.1016/j.jplph.2009.01.002
- Lopez-Serra, P., Marcilla, M., Villanueva, A., Ramos-Fernandez, A., Palau, A., Leal, L., et al. (2014). A DERL3-associated defect in the degradation of SLC2A1 mediates the warburg effect. *Nat. Commun.* 5, 3608. doi: 10.1038/ncomms4608
- Martínez-Prada, M. M., Curtin, S. J., and Gutiérrez-González, J. J. (2021). Potato improvement through genetic engineering. *GM Crops Food* 12 (1), 479–496. doi: 10.1080/21645698.2021.1993688
- McKibbin, R. S., Muttucumar, N., Paul, M. J., Powers, S. J., Burrell, M. M., Coates, S., et al. (2006). Production of high-starch, low-glucose potatoes through over-expression of the metabolic regulator SnRK1. *Plant Biotech. J.* 4, 409–418. doi: 10.1111/j.1467-7652.2006.00190.x
- Moorhead, G., Douglas, P., Cotellet, V., Harthill, J., Morrice, N., Meek, S., et al. (1999). Phosphorylation-dependent interactions between enzymes of plant metabolism and 14-3-3 proteins. *Plant J.* 18 (1), 1–12. doi: 10.1046/j.1365-313x.1999.00417.x
- Naess, S., Bradeen, J., Wielgus, S., Haberlach, G. T., McGrath, J. M., and Helgersson, J. P. (2000). Resistance to late blight in solanum bulbocastanum is mapped to chromosome 8. *Theor. Appl. Genet.* 101, 697–704. doi: 10.1007/s001220051533
- Nakaminami, K., Matsui, A., Nakagami, H., Minami, A., Nomura, Y., Tanaka, M., et al. (2014). Analysis of differential expression patterns of mRNA and protein during cold-acclimation and de-acclimation in arabidopsis. *Mol. Cell Proteomics* 13, 3602–3611. doi: 10.1074/mcp.M114.039081
- Noguero, M., Atif, R. M., Ochatt, S., and Thompson, R. D. (2013). The role of the DNA-binding one zinc finger (DOF) transcription factor family in plants. *Plant Sci.* 209, 32–45. doi: 10.1016/j.plantsci.2013.03.016
- Oecking, C., Eckerskorn, C., and Weiler, E. W. (1994). The fusicoccin receptor of plants is a member of the 14-3-3 superfamily of eukaryotic regulatory proteins. *FEBS Lett.* 352 (2), 163–166. doi: 10.1016/0014-5793(94)00949-x
- Oñate-Sánchez, L., and Vicente-Carbajosa, J. (2008). DNA-Free RNA isolation protocols for arabidopsis thaliana, including seeds and siliques. *BMC. Res. Notes* 1, 93. doi: 10.1186/1756-0500-1-93
- Park, D. H., Somers, D. E., Kim, Y. S., Choy, Y. H., Lim, H. K., Soh, M. S., et al. (1999). Control of circadian rhythms and photoperiodic flowering by the arabidopsis GIGANTEA gene. *Science* 285, 1579–1582. doi: 10.1126/science.285.5433.1579
- Patil, V. U., Singh, R., Vanishree, G., Duti, S., Kavar, P. G., Bhardwaj, V., et al. (2016). Genetic engineering for enhanced nutritional quality in potato—a review. *Potato J.* 43 (1), 1–21.
- Peña, P. A., Quach, T., Sato, S., Ge, Z., Nersesian, N., Changa, T., et al. (2017). Expression of the maize Dof1 transcription factor in wheat and sorghum. *Front. Plant Sci.* 8. doi: 10.3389/fpls.2017.00434
- Pourazari, F., Andersson, M., and Weih, M. (2018). Altered tuber yield in genetically modified high-amylose and oil potato lines is associated with changed whole-plant nitrogen economy. *Front. Plant Sci.* 9. doi: 10.3389/fpls.2018.00342
- Prescha, A., Swiedrych, A., Biernat, J., and Szopa, J. (2001). Increase in lipid content in potato tubers modified by 14-3-3 gene overexpression. *J. Agric. Food Chem.* 49, 3638–3643. doi: 10.1021/jf010258o
- Queval, G., and Noctor, G. (2007). A plate reader method for the measurement of NAD, NADP, glutathione, and ascorbate in tissue extracts: Application to redox profiling during arabidopsis rosette development. *Anal. Biochem.* 363 (1), 58–69. doi: 10.1016/j.ab.2007.01.005
- Ralet, M. C., and Gueguen, J. (2001). Foaming properties of potato raw proteins and isolated fractions *Lebensmittel-wissenschaft und technologie* 34, 266–269. doi: 10.1006/ftsl.2000.0684
- Ramírez-Gonzales, L., Shi, L., Bergonzi, S. B., Oortwijn, M., Franco-Zorrilla, J. M., Solano-Tavira, R., et al. (2021). Potato CYCLING DOF FACTOR 1 and its lncRNA counterpart StFLORE link tuber development and drought response. *Plant J.* 105 (4), 855–869. doi: 10.1111/tj.15093
- Ramos-Fernández, A., Paradelo, A., Navajas, R., and Albar, J. P. (2008). Generalized method for probability-based peptide and protein identification from tandem mass

- spectrometry data and sequence database searching. *Mol. Cell Proteomics* 7 (9), 1748–1754. doi: 10.1074/mcp.M800122-MCP200
- Reeds, P. J. (2000). Dispensable and indispensable amino acids for humans. *J. Nutr.* 130 (7), 1835S–1840S. doi: 10.1093/jn/130.7.1835S
- Regierer, B., Fernie, A. R., Springer, F., Perez-Melis, A., Leisse, A., Koehl, K., et al. (2002). Starch content and yield increase as a result of altering adenylate pools in transgenic plants. *Nat. Biotechnol.* 20, 1256–1260. doi: 10.1038/nbt760
- Reimand, J., Arak, T., Adler, P., Kolberg, L., Reisberg, S., Peterson, H., et al. (2016). g:Profiler a web server for functional interpretation of gene lists, (2016 Update). *Nucl. Acids Res.* 44 (8), 83–89. doi: 10.1093/nar/gkw199
- Renau-Morata, B., Carrillo, L., Cebolla-Cornejo, J., Molina, R. V., Martí, R., Domínguez-Figueroa, J., et al. (2020b). The targeted overexpression of SICDF4 in the fruit enhances tomato size and yield involving gibberellin signalling. *Sci. Rep.* 10, 10645. doi: 10.1038/s41598-020-67537-x
- Renau-Morata, B., Carrillo, L., Domínguez-Figueroa, J., Vicente-Carbajosa, J., Molina, R. V., Nebauer, S. G., et al. (2020a). CDF transcription factors: plant regulators to deal with extreme environmental conditions. *J. Exp. Bot.* 71 (13), 3803–3815. doi: 10.1093/jxb/eraa088
- Renau-Morata, B., Molina, R. V., Carrillo, L., Cebolla-Cornejo, J., Sánchez-Perales, M., Pollmann, S., et al. (2017). Ectopic expression of CDF3 genes in tomato enhances biomass production and yield under salinity stress conditions. *Plant Sci.* 8. doi: 10.3389/fpls.2017.00660
- Roessner, U., Willmitzer, L., and Fernie, A. R. (2001). High-resolution metabolic phenotyping of genetically and environmentally diverse potato tuber systems: identification of phenocopies. *Plant Physiol.* 127, 749–764. doi: 10.1104/pp.010316
- Roessner-Tunali, U., Urbanczyk-Wochniak, E., Czechowski, T., Kolbe, A., Willmitzer, L., and Fernie, A. R. (2003). *De novo* amino acid biosynthesis in potato tubers is regulated by sucrose levels. *Plant Physiol.* 133 (2), 683–692. doi: 10.1104/pp.103.024802
- Rueda-Lopez, M., Pascual, M. B., Pallero, M., Henao, L. M., Lasa, B., Jauregui, I., et al. (2017). Overexpression of a pine dof transcription factor in hybrid poplars: a comparative study in trees growing under controlled and natural conditions. *PLoS One* 12, e0174748. doi: 10.1371/journal.pone.0174748
- Santelia, D., and Zeeman, S. C. (2010). Progress in arabidopsis starch research and potential biotechnological applications. *Curr. Opin. Biotech.* 22, 271–280. doi: 10.1016/j.copbio.2010.11.014
- Sawa, M., Nusinow, D. A., Kay, S. A., and Imaizumi, T. (2007). FKF1 and GIGANTEA complex formation is required for day-length measurement in arabidopsis. *Science* 318, 261–265. doi: 10.1126/science.1146994
- Schwacke, R., Ponce-Soto, G. Y., Krause, K., Bolger, A. M., Arsova, B., Hallab, A., et al. (2019). MapMan4: A refined protein classification and annotation framework applicable to multi-omics data analysis. *Mol. Plant* 12, 879–892. doi: 10.1016/j.molp.2019.01.003
- Schwender, J., König, C., Klapperstück, M., Heinzel, N., Munz, E., Hebbelmann, I., et al. (2014). Transcript abundance on its own cannot be used to infer fluxes in central metabolism. *Front. Plant Sci.* 5. doi: 10.3389/fpls.2014.00668
- Sehnke, P. C., Chung, H. J., Wu, K., and Ferl, R. J. (2001). Regulation of starch accumulation by granule-associated plant 14-3-3 proteins. *Proc. Natl. Acad. Sci. U.S.A.* 98, 765–770. doi: 10.1073/pnas.98.2.765
- Singh, R. P., and Anderson, B. A. (2004). *Understanding and measuring the shelf-life of food*. Ed. R. Steele, ISBN: . Woodhead Publishing limited. CRC Press. Cambridge, England, chapter 1, pp 3-19
- Smith, E. N., Schwarzländer, M., Ratcliffe, R. G., and Kruger, N. J. (2021). Shining a light on NAD- and NADP-based metabolism in plants. *Trends Plant Sci.* 10, 1072–1086. doi: 10.1016/j.tplants.2021.06.010
- Swiedrych, A., Prescha, A., Matysiak-Kata, I., Biernat, J., and Szopa, J. (2002). Repression of the 14-3-3 gene affects the amino acid and mineral composition of potato tubers. *J. Agric. Food Chem.* 50, 2137–2141. doi: 10.1021/jf0112825
- Tarn, T. R., Tai, G. C. C., and Liu, Q. (2006). “Chapter 5: Quality improvement,” in *Hand-book of potato production, improvement, and postharvest management*. Eds. J. Gopal and S. M. P. Khurana (New York: Haworth Press), 147–178.
- Tek, A. L., Stevenson, W. R., Helgeson, J. P., and Jiang, J. (2004). Transfer of tuber soft rot and early blight resistances from *Solanum tuberosum* to cultivated potato. *Theor. Appl. Genet.* 109, 249–254. doi: 10.1007/s00122-004-1638-4
- Tjaden, J., Möhlmann, T., Kampfenkel, K., Henrichs, G., and Neuhaus, H. E. (1998). Altered plastidic ATP/ADP-transporter activity influences potato (*Solanum tuberosum* L.) tuber morphology, yield and composition of tuber starch. *Plant J.* 16, 531–540. doi: 10.1046/j.1365-313x.1998.00317.x
- Toinga-Villafuerte, S., Vales, M. I., Awika, J. M., and Rathore, K. S. (2022). CRISPR/Cas9-mediated mutagenesis of the granule-bound starch synthase gene in the potato variety Yukon gold to obtain amylose-free starch in tubers. *Int. J. Mol. Sci.* 23, 4640. doi: 10.3390/ijms23094640
- Toroser, D., Athwal, G. S., and Huber, S. C. (1998). Site-specific regulatory interaction between spinach leaf sucrose-phosphate synthase and 14-3-3 proteins. *FEBS Lett.* 435, 0014–5793. doi: 10.1016/S0014-5793(98)01048-5
- van Dongen, J. T., Gupta, K. J., Ramírez-Aguilar, S. J., Araújo, W. L., Nunes-Nesi, A., and Fernie, A. R. (2011). Regulation of respiration in plants: a role for alternative metabolic pathways. *J. Plant Physiol.* 168, 1434–1443. doi: 10.1016/j.jplph.2010.11.004
- Van Koningsveld, G. A., Waltra, P., Voragen, A. G., Kuijpers, I. J., van Boekel, M. A., and Gruppen, H. (2006). Effect of protein composition and enzymatic activity on formation and properties of potato protein stabilized emulsions. *J. Agric. Food Chem.* 54, 6419–6427. doi: 10.1021/jf061278z
- Varala, K., Marshall-Colón, A., Cirrone, J., Brooks, M. D., Pasquino, A. V., Lérán, S., et al. (2018). Temporal transcriptional logic of dynamic regulatory networks underlying nitrogen signalling and use in plants. *Proc. Natl. Acad. Sci. U.S.A.* 115, 6494–6499. doi: 10.1073/pnas.1721487115
- Wurzinger, B., Nukarinen, E., Nägele, T., Weckwerth, W., and Teige, M. (2018). The SnRK1 kinase as central mediator of energy signaling between different organelles. *Plant Physiol.* 176 (2), 1085–1094. doi: 10.1104/pp.17.01404
- Yanagisawa, S. (2002). The dof family of plant transcription factors. *Trends Plant Sci.* 7, 555–560. doi: 10.1016/S1360-1385(02)02362-2
- Yanagisawa, S. (2004). Dof domain proteins: plant-specific transcription factors associated with diverse phenomena unique to plants. *Plant Cell Physiol.* 45, 386–391. doi: 10.1093/pcp/pch055
- Yanagisawa, S. (2016). “Structure, function and evolution of the dof transcription factor family,” in *Plant transcription factors* (New York, NY, USA: Elsevier), 183–197.
- Yanagisawa, S., and Schmidt, R. J. (1999). Diversity and similarity among recognition sequences of dof transcription factors. *Plant J.* 17, 209–214. doi: 10.1046/j.1365-313x.1999.00363.x
- Yanagisawa, S., and Sheen, J. (1998). Involvement of maize dof zinc finger proteins in tissue-specific and light-regulated gene expression. *Plant Cell.* 10, 75–89. doi: 10.1105/tpc.10.1.75
- Zuk, M., Weber, R., and Szopa, J. (2005). 14-3-3 protein down-regulates key enzyme activities of nitrate and carbohydrate metabolism in potato plants. *J. Agric. Food Chem.* 53 (9), 3454–3460. doi: 10.1021/jf0485584